SYNTHESIS OF $(-)$ -TALAROMYCINS A AND B^{\dagger}

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Abstract -- Highly enantiomerically pure (-)-talaromycins A and B [(3R, 4S, 6R, 9R)- and (3S,4S,6R,9R)-9-ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro(5.5)undecane] were synthesized starting from chiral building blocks of microbial origin.

(-)-Talaromycins A 1a and B 2a were fungal toxins isolated by Lynn and co-workers in 1982 from Talaromyces stipitatus.^{1,2} The toxicity of these spiroacetals may be due to their ability to block outward potassium fluxes, thus leading to muscle dysfunction.¹ Their unique spiroacetal structures la and 2a attracted much attention of synthetic chemists. Thus thermodynamically more stable (±)-talaromycin B 2a with an eq CH₂OH group was synthesized by Schreiber et al.,³ Kozikowski et al.,⁴ Kocienski et al.⁵ and Kay et al.⁶ The less stable (±)-talaromycin A 1a with an ax CH₂OH group was later synthesized by Schreiber et $a1.^7$ As to the synthesis of the naturally occurring enantiomers of talaromycins, there exist only two reports.^{8,9} Smith and Thompson was the first to synthesize (-)-talaromycins A and B in 1984.⁸ The second synthesis of (-)-talaromycin A was reported by Midland and Gabriel in 1985.⁹ As an extension of our continuing efforts to synthesize enantiomerically pure spiroacetals of insect origin,^{10,11} we initiated a project to synthesize both (-)-talaromycins A and B in highly enantiomerically pure state.

Our synthetic plan is shown in Fig. 1. Chiral building blocks of microbial origin are employed as our starting materials. For the construction of the spiroacetal system, a Wittig reaction^{12~14} between A and B is to be employed. The phosphorane A can be prepared from C, which in turn is the product of yeast reduction^{15~17} of D. Dimethyl 3-oxopentanedioate E is the starting material for D. The aldehyde B is to be synthesized via F from ethyl (S)-3-hydroxybutanoate G, the product of yeast reduction of ethyl acetoacetate.¹⁸ Execution of the above plan, which culminated to the synthesis of crystalline (-)-talaromycin A 1a, will be detailed below.

The first phase of our work as shown in Fig. 2 was the synthesis of a phosphonium salt 14, the precursor to the Wittig reagent A. Reduction of 3 (=E) with NaBH₄ gave 4a. After protecting the OH group of 4a as a THP ether, the resulting 4b was reduced with LAH

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Tsynthetic Microbial Chemistry - XIII. Part XII, T. Kitahara, H. Kurata, T. Matsuoka and K. Mori, T<u>etrahedron</u> 41, 5475 (1985). The experimental part of this work was taken from the forthcoming doctoral dissertation of M. I. (March, 1987).

Pig. 1. Synthetic plan.

to give **5a.** This was treated with p-TsOH *in* MeOH to give a trio1 **5b.** Acetonide 6 was prepared from **5b** in the usual manner. Oxidation of 6 with pyridinium chlorochromate (PCC)¹⁹ in the presence of MS $3\overset{20}{h}20$ furnished aldehyde 7. When a soln of 7 and p-TsOH in CH_2Cl_2 -EtOH was heated, intramolecular acetalization took place to give a stereoisomeric mixture of 8 in 64 % yield from 7 or in 33 % overall yield from 3 in 7 steps. Although a synthesis of 4-hydroxy-2-methoxytetrahydropyran was known,²¹ the present multi-step route furnished 8 in better overall yield. The alcohol 8 was oxidized with pyridinium dichromate (PDC)²² in the presence of MS $3A^{20}$ to give 9 in 72 % yield. Ethoxycarbonylation of 9 could not be achieved under the conventional condition employing NaH and CO(OEt)₂. By the method of Mander and Sethi, ²³ however, a CO₂Et group was successfuly introduced at C-5 of 9. Thus 9 was treated with LDA and NCCO₂Et²⁴ to give a mixture of two regioisomeric 8-keto esters 10 and 11 in a ratio of **1O:l.** This was purified by Si02 chromatography to give pure **10** in 55 % yield.

The next step was the crucial microbial reduction of **10.** Reduction of the 8-keto ester 10 was tried with Saccharomyces bailii KI 0116, Pichia terricola KI 0117 and Saccharomyces cerevisiae (baker's yeast).¹⁸ S. bailii and P. terricola were found to be unsuitable for this reduction giving almost no β -hydroxy ester, although \underline{S} . bailii was an excellent organism to reduce a β -keto ester related to PGI₂.²⁵ Reduction of 10 was therefore carried out with baker's yeast. An emulsion of **10** in dilute Triton X-100 was added to a suspension of briskly fermenting baker's yeast at pH 8 (phosphate buffer) in the presence of sucrose. The product obtained in 81 % yield was a diastereomeric mixture at C-2 of hydroxy ester **12a.** The diastereomeric ratio was later determined as 63:37 by analyzing 13c (vide infra). The diastereomers of 12a were partially separated by SiO₂ chromatography to give the less polar isomer (major product), the more polar isomer (minor product) and their mixture. In the NMR spectrum of the less polar isomer of 12a, a signal

Pig. 2. **Synthesis of the phosphonium salt Ii.**

due to an eq H at C-2 (δ 4.78, 1H, dd, \underline{J} =3 and 3 Hz) and that due to an ax H at C-5 (δ 2.53, 1H, ddd, $J=10$, 4 and 2 Hz) were observed. In all of the published examples of the yeast reduction of cyclic B-keto esters, formation of cis-B-hydroxy esters were observed with (S)-configuration of the OH group.^{15~17,25,26} The less polar isomer of 12a was therefore assumed to be $(2S, 4S, 5R)$ -12a, whose enantiomeric purity was estimated to be 43 % e.e. by the HPLC analysis of the corresponding (\underline{R}) - and (\underline{S}) - α -methoxy- α -trifluoromethylphenylacetates (MTPA esters²⁷), (2<u>5,45,5R</u>)-12b and 12c. The more polar and minor isomer of 12a exhibited in its NMR spectrum a signal due to an eq H at C-2 (6 4.71, 1H, dd, J=4.4 and 3.6 Hz) and that due to an eq H at C-5 (δ 2.62, 1H, ddd, $J=4.4$, 4 and 3.2 Hz). Assuming the (S)-configuration for 4-OH, this isomer was thought to be $(2g,4S,5R)$ -12a. Determination of the enantiomeric purity of $(2R, 4S, 5R) - 12a$ was rather troublesome, because its MTPA ester (2R, 4S, 5R)-12b showed no good separation of the diastereomers when analyzed by HPLC. The minor isomer $(2R, 4S, 5R)$ -12a was therefore treated with p-TsOH in EtOH to effect equilibration at C-2 to generate $(2S, 4S, 5R)$ -12a, whose MTPA ester $(2S, 4S, 5R)$ -12b was known to exhibit good diastereomer separation by HPLC. The HPLC analysis of the MTPA esters derived from the above described equilibrated mixture showed the generated (2&4S,SIJ)-12a to be **of 100 % e.e.** Accordingly the diastereomeric mixture of 12a was of 64 % e.e. [(43 x 0.63) + (100 x 0.37) = 64] with regard to the chiral centers at C-4 and c-5.

To secure a supporting evidence for the assumed absolute configuration of the two isomers of 12a, they were converted to the corresponding dibenzoates 13c and their CD spectra were measured. To prepare 13c, 12a (diastereomeric mixture) was first acetylated with Ac₂0 to **12d.** LAH reduction of **12d** gave **13a** in 97 % yield from **12a.** Direct reduction of 12a with LAH afforded an inferior result. Benzoylation of 13a yielded dibenzoates 13c as a diastereomeric mixture at C-2 in a ratio of 63:37 as analyzed by HPLC. The diastereomers of 13c were separated by prep TLC to give the less polar isomer (21 % recovery) and the more polar isomer (52 % recovery). 1_H NMR spectra of these two isomers revealed that the former possessed an ax H at C-4 (δ 5.70, 1H, ddd, J=7.3, 4.4 and 4.1 Hz), while the latter had an eq H at C-4 (6 5.47, 1H, ddd, $J=5.8$, 5.8 and 5.8 Hz). In both of them the C-2 H was in eq orientation $(\underline{J}=3.3-4.9$ Hz) like in the starting β -hydroxy esters 12a. Application of the exciton chirality method^{28,29} to these dibenzoate isomers provided a supporting evidence concerning their absolute configuration. The exciton-split CD spectrum of the less polar isomer of 13c in EtOH showed a positive first Cotton effect at 237 nm (AE **+ll)** and a negative second Cotton effect at 220 nm (As -2.8). The more polar isomer of 13c also exhibited a positive first Cotton effect at 238 nm ($\Delta \epsilon$ +5.8) and a negative second Coton effect **at** 220 nm (AE -1.0). Comparison of these data with the CD spectra of some triterpene dibenzoates of similar structural feature²⁸ supported the assumed (4S)-configuration of the dibenzoates 13c as depicted in Fig. 3. The less polar isomer was therefore $(2R, 4S, 5S)$ -13c and the more polar one was $(2S, 4S, 5S)$ -13c.

Pig. 3. CD spectra of (2R,4S,5S)-13c and (2S,4S,5S)-13c.

Having clarified the absolute configuration and enantiomeric purity of **13a,** the remaining task was its conversion to 14. Treatment of a diastereomeric mixture of 13a with PhCH₂Br and NaH in the presence of $(p_f$ -Bu)₄NI in THF^{Cf.30} gave the corresponding dibenzyl ether 13b in 80 % yield. The desired phosphonium salt 14 was prepared in guantitative yield from 13b by heating it with $Ph_3PH^+BF_4^-$ in MeCN.^{31,32} The overall yield of 14 from 3 was 8 % in 13 steps.

The second phase of our project as shown in Fig. 4 was the synthesis of the aldehyde 20 (=B), the other partner of the Wittig reaction. Reduction of ethyl acetoacetate 15 with a thermophilic yeast Saccharomyces bailii KI 0116 gave (S)-16 (96~98 % e.e.) in 84 % yield.¹⁸ The dianion derived from 16 was alkylated with allyl bromide according to Frater to give 17a contaminated with its syn-isomer (anti:syn=96:4) in 85 % yield.³³ The diastereomeric ratio was determined by capillary GLC using a diastereomeric mixture of ethyl

Pig. 4. **8ynthesls of the aldehyde 2g.**

(t)-2-(1'-hydroxyethyl)pent-4-enoate (anti:syn=51:49) as a reference sample. Conversion of 17a to 28 was lengthy but straightforward. The OH group of 17a was protected as THP ether. Reduction of the resulting **17b** with LAH gave an alcohol 18a. This was benzylated to 18b, and the THP protective group was removed from 18b to give 18c. The corresponding 3,5_dinitrobenzoate (DNB) 18d was crystalline, and could be purified by recrystallization to give diastereomerically and enantiomerically pure **led.** Treatment of **18d** with KOH gave pure 18c, whose diastereomeric purity as 100 % was checked by capillary GLC. The corresponding (\underline{R}) -MTPA ester 18e exhibited a single peak upon HPLC analysis proving the high enantiomeric purity (100 $%$ e.e.) of 18 c . As the task of the OH group to control the absolute configuration **of** the new chiral center was over, it was removed by tosylation to 18f followed by its reduction with LiBEt₃H.³⁴ The benzyl (Bn) protective group of 19a was then replaced by the THP group to give 19c by reducing 19a with Na/liq NH₃ and protecting the OH group of **19b.** Finally, the Lemieux-Johnson oxidation of 19c with Os04-NaI04 gave (R)-20 in 34 % overall yield from 15 in 13 steps. By switching the protective group from Bn to THP, the construction of the spiroacetal system became less complicated due to the more facile deprotection of the THP group compared with that of the Bn group.

The final stage of the synthesis as shown in Fig. 5 was the coupling of the two building blocks 14 and 20 by the Wittig reaction to give the desired 1,7-dioxaspiro[5.5] undecane system. Treatment of the phosphonium salt 14 with n-BuLi in THF-HMPA generated a deep red phosphorane 21. This was condensed with the aldehyde 20 to give a cyclic enol ether 22. Acid treatment (conc HCl:H₂O:THF=1:5:20) of crude 22 gave a mixture of spiroacetals. As one of the building blocks was optically impure (64 % e.e.), formation of four spiroacetal dibenzyl ethers **(lb, 23b, 25b** and **26b)** were expected. However, the products obtained after SiO₂ chromatography were a mixture of dibenzyl ethers 1b and 23b (46 $%$ from 14) and also a mixture of 12-monobenzyl ethers 1c, 2c and 24c (35 $%$ from 14). The unexpected formation of 12-monobenzyl ethers could be rationalized as shown in Fig. 5 by the sequence $22 \div \alpha \div \beta + \gamma + 1c + 2c + 24c$ involving (i) olefin isomerization (ii) retro-Michael-like elimination of PhCH₂OH (iii) addition of H₂O and (iv) spiroacetal formation. Under the acid condition employed, the above process competed to some extent with the desired hydrolysis of the THP group followed by spiroacetalization. The elimination-addition mechanism as above allowed the formation **of** 2c and Zlc, where the OH group and the CH₂OBn group were in trans-relationship in contrast to the original cis-relation-

Synthesis of Talaromycins A and B. Fig. 5.

ship in 21. The structures 1b, 23b, 1c, 2c and 24c were deduced on the basis of the NMR analysis of thir 4-benzoates (1f, 2f and 24f) or that of their 12-mono DNB derivatives 1e, 2e and 23e (see Experimental). Other two possible isomers could not be isolated from the reaction mixture.

Crystalline 12-mono DNB derivative le of $(3R, 4S, 6R, 9R)$ -talaromycin A la was derived from the mixture of the dibenzyl ethers lb and 23b in the following manner. The mixture $(2.35 g)$ of **1b** and 23b was reduced with Na/liq NH₃ to give a mixture of 1a and 23a in 75 % yield. Treatment **of** the mixture of la and 23a with 3,5-dinitrobenzoyl chloride (DNBCl) in pyridine in the presence of $4-(N,N-dimethy)$ amino)pyridine (DMAP) gave a complex mixture of products. This was chromatographed over $SiO₂$ to give, in the order of elution, a mixture of bis LWB esters **(Id** and 23d; 18 % yield), a mixture of new mono DNB esters (25e and 26e; 14 % yield), and a mixture of talaromycin A 12-mono DNB ester le and its isomer 23e (40 % yield). The new mono DNB esters 25e and 26e must have been generated by epimerization at the spiro-center in the course of the acylation reaction, because they could be detected by TLC monitoring of the acylation reaction. The epimerization was due to the preference of the CH₂ODNB group to adopt eq orientation. Pure talaromycin A 12-mono DNB ester 1e (509 mg), m.p. 147-148°, $[\alpha]_D^{18}$ -113° (CHCl₃), was obtained by recrystallization of the mixture (730 mg) of 1e and 23e. Pure 23e (123 mg), m.p. 171.5~173°, [a] $_{10}^{18}$ +136° (CHCl₃), was also secured by purification with prep TLC and recrystallization. The mixture of bis DNB esters (1d and 23d) was hydrolyzed with K_2CO_3 to a mixture of 1a and 23a, which was acylated again and recrystallized to give additional amounts of le (62 mg) and 23e (26 mg). A further amount of **le (93** mg) was obtained from a mixture **(1.10 g)** of monobenzyl ethers (1c, 2c and 24c) by reduction with Na/liq NH₃, acylation with DNBCl and recrystallization of the resulting mixture of mono DNB esters **le, 2e** and 24e. In total, talaromytin A 12-mono DNB ester **le was** obtained in 13 % overall yield from **13b** in 5 steps. From the combined mother liquor of the above-mentioned recrystallization experiments, small amounts of pure talaromycin B 12-mono DNB ester 2e, m.p. 104~104.2°, [α] $^{24}_{\rm D}$ -29.8° (CHCl₃), and another mono DNB ester 25e, m.p. 128.8~129.2°, [$\alpha1\frac{24}{D}$ -77.7° (CHCl₃), were isolated. The amount of pure 2e was so small that its hydrolysis to talaromycin B 2a was not attempted. In Fig. 6 is shown the ratio of four 12-mono LXiB esters **(le, 23e, 25e** and 26e) generated from the mixture of dibenzylethers **(lb** and **23b) as** expressed in percentages. As can be seen from Fig. 6, the ratio of the isomers reflected the enantiomeric purity of the phosphorane 21.

Fig. 6. Derivation of the 12- mono DNB esters of talaromycin stereoisomers.

To complete the synthesis, talaromycin A mono DNB ester 1e was hydrolyzed with $K_2\omega_3$ in MeOH-THF to give,in 89 % yield, the natural enantiomer of talaromycin A la for the first time as crystals, m.p. $19-20^{\circ}$, $[\alpha]_D^{24}$ -146° (CHCl₃) [lit.⁸ $[\alpha]_D^{20}$ -110.2° (CHCl₃);

lit.⁹ [α] $^{26}_{16}$ -124.9° (CHCl₃)]. Our success in synthesizing highly pure and crystalline f-)-talaromycin A la may be due to the nicely crystalline nature of its t2-mono DNB ester le. Because of that property we were able to purify it completely. (-)-Talaromycin B Za, $\lceil \alpha \rceil_0^{24}$ -89.1° (CHCl₃) [lit.⁸ $\lceil \alpha \rceil_0^{20}$ -84.1° (CHCl₃)], was also prepared from 1e by hydrolysis with K_2CO_3 followed by acid-catalyzed isomerization with Amberlyst[®]-15 (H⁺-form) in MeOH in 78 % yield. The 400 MHz 1 H NMR spectra of our synthetic (-)-talaromycins A and B were in good accord with the authentic spectra of the natural products kindly provided by Prof. Lynn (360 MHz)¹ and Prof. Smith (250 and 500 MHz).⁸ The ¹³C NMR spectrum of (-)-talaromycin B was in accord with the data published for (t) -talaromycin B.⁵ The CD spectra of the dibenzoates 1g and 2g of (-)-talaromycins A and B are shown in Fig. 6. They were also

Fig. 7. **CD spectra of the dibenroates of talarcmycins A and B.**

identical **to** Prof. Lynn's authentic spectra of the dibenzoates derived from the natural products. The overall yield of **la** was 4 % from ethyl acetoacetate 15 in 18 steps or 0.9 % from dimethyl 3-oxopentanedioate 3 in 19 steps. $(-)$ -Talaromycin B 2a was obtained in 3 % overall yield from **15** in 19 steps or in 0.8 % overall yield from 3 in 20 steps.

In summary, highly pure natural enantiomers of talaromycins A and B were synthesized in a convergent manner employing the chiral building blocks of microbial origin. The yeast reduction was again shown to be useful in organic synthesis. An enantiomerically impure intermediate such as 21 can satisfactorily be employed to afford enantiomerically pure target molecules after removing the unwanted diastereomers. Experimental importance of a crystalline intermediate in a chiral synthesis was also exemplified in the present case. The principle of 'optical enrichment'^{9,11} in the case of a chiral synthesis was applicable to the present synthesis, too.

EXPERIMENTAL

All bes and mes were uncorrected. IR spectra were measured as films for oils or as mujol mulls or KBr discs for

solids on a Jasco IRA-102 or IRA-202 spectrometer unless otherwise stated. ¹H NMR spectra were recorded with TMS as an internal standard on a Hitachi R-24A (60 MHz) or on a JEOL JNM FX-100 (100 MHz, CDCl₃) or on a JEOL JNM GX-400 (400 MHz, CDCl₃) spectrometer, ¹³C NMR spectra were measured with TMS as an internal standard as CDCl₃ soln at 25 MHz on a Jeol JNM FX-100 spectrometer. Optical rotations were measured on a Jasco DIP 140 polarimeter as CHCl₃ soln. ORD and CD spectra were measured on a Jasco J-20C spectropolarimeter. Mass spectra were recorded on a JBOL DX-303 or on a Hitachi RMU-6M spectrometer at 70 eV. Puji-Davison BW-820 MH was used for SiO₂ column chromatography. Merck Kieselgel 60 Art 5717 or 5744 were used for prep TLC separation. HPLC analyses were performed on Nucleosil® 50-5 (25 cm x 4.6 mm) by the detection at 254 nm unless otherwise stated.

Dimethyl 3-tetrahydropyranyloxypentanedicate 4b. Dimethyl 3-oxopentanedicate 3 (200 g, 1.15 mol) was reduced with NaBHA (16.7 g, 0.44 mol) in MeCH (700 ml) in a usual manner to give crude dimethyl 3-hydroxypentanedicate 4a (213 g, quantitative), whax 3550 (s), 1740 (s), 1200 (s) cm⁻¹. Crude 4a (210.3 g, ~1.14 mol) was treated with dihydropyran (138 g, 1.64 mol) and p-TsOH-H₂O (0.4 g, 2.1 mmol) in Et₇O (1.2 l) in a usual manner to give crude 4b (296.9 g, quantitative). Analytical sample: b.p. 105~121°/1.0 Torr; n¹⁶₁5</sup> 1.4521; vmax 1740 (s), 1200 (s), 1030 (s) cm⁻¹; 2.47, 2.54, 2.57 and 2.62 (total 4H, each d, J=6 Hz), 3.10~3.90 (2H, m), 3.64 (6H, s), 4.36 (1H, t, J=6 Hz), 4.72 (1H, br.s). (Found: C, 55.47; H, 7.68. Calc for C₁₂H₂₀O₆: C, 55.37; H, 7.75 %).

Pentane-1,3,5-triol 3-THP ether 5a. Crude 4b (129 g, 0.50 mol) was reduced with LiAlH4 (LAH, 30 g, 0.69 mol) in Et₂0 (720 ml) in a usual manner to give crude 5a (100 g, 98 % from 3), vmax 3420 (s), 1140 (s), 1080 (s), 1030 (s) cm⁻¹; 8 (CDCl₃) 1.00~2.40 (10H, m),3.32 (2H, s, OHx2), 3.10~4.38 (7H, m), 4.4~4.8 (1H, br.s).

Pentane-1,3,5-triol 5b. Crude 5a (235.7 g, ~1.15 mol) was treated with p-TsOH.H₂O (2.6 g, 14 mmol) in MeOH (1.2 l) in a usual manner to give crude 5b (146,9 g, quantitative). Analytical sample: b.p. 154~156°/1.3 Torr; n¹⁸ 1.4655; vmax 3400 (s), 1050 (s) cm^{-1} ; 6 (C₅D₅N-CDCl₃) 2.00 (4H, dt, J=6, 6 Hz), 4.09 (4H, t, J=6 Hz), 4.43 (1H, t, J=6 Hz), 5,86 (3H, s, OHx3), (Found: C, 50,35; H,10,15, Calc for C₅H₁₂O₃: C, 49,98; H, 10.07 a).

Pentane-1,3,5-triol 1,3-acetonide 6. Crude 5b (144 g, 1.13 mol) was treated with 2,2-dimethoxypropane (254 g, 2.4 mol) and p-TBOH-H₂O (6.5 g, 34 mmol) in acetone (1.2 1) in a usual manner to give 6 (98.3 g, b.p. 60-107°/15-16 Torr, 53 % from 3). A portion was chromatographed over SiO_2 (n-hexane-Et₂O (15:1~2:1)] followed by distillation in the presence of $K_2\text{CO}_3$ to give an analytical sample: b.p. 113-115°/17 Torr; n⁸ 1.4495; vmax 3450 (s), 1200 (s), 0.95~1.45 (2H, m), 1.32 (3H, s), 1.43 (3H, s), 1.63 (2H, dt, J=5, 5 Hz), 3.06 (1H, t, J=5 Hz, OH), 3.25~4.31 (4H, m). (Found: C, 60.40; H, 10.05. Calc for CgH₁₆03: C, 59.98; H, 10.07 %).

3.5-Dihydroxypentanal acetonide 7. According to the reported procedure, 19,20 6 (20 g, 125 mmol) was oxidized with pyridinium chlorochromate (PCC, 45.4 g, 211 mmol) in CH₂Cl₂ (310 ml) in the presence of MS 3Å (40 g) to give crude 7 (19.3 g, 97 W. A portion of it was chromatographed over SiO_2 [n-pentane-Et₂0 (20:1-3:1)] followed by distillation under Ar to give an analytical sample; b.p. 83-87°/45 Torr; n¹⁸ 1.4419; wmax 2750 (m), 1730 (s), 1200 (s), 1165 (s), 1100 (s) cm⁻¹; 6 (CCl₄) 1.26 (3H, s), 1.41 (3H, s), 0.80~1.90 (2H, m), 2.45 (2H, ddd, J=2, 2, 6 Hz), 2.46 (1H, dd, J=2, 6 Hz), 3.40~4.00 (2H, m), 4.00~4.63 (1H, m), 9.86 (1H, dd, J=2, 2 Hz). (Found: C, 60.31; H, 8.87. Calc for C_BH₁₄0₃: C, 60.74; H, 8.92 %).

2-Ethoxytetrahydropyran-4-ol 8 A soln of crude 7 (18.2 g, ~115 mmol) and p-TsOH·H₂O (0.4 g, 2.1 mmol) in 99 & EtOH (80 ml) and CH₂Cl₂ (240 ml) was stirred and heated under reflux overnight with azeotropic removal of water by use of MS 4A. After cooling, the mixture was neutralized by the addition of solid Na₂O₃ (4.6 g, 43.4 mmol) and stirred further for 4 h at room temp. The solid was filtered off through a pad of Celite and washed with Et₂O. The combined filtrate and washings were concentrated in vacuo. The residue was diluted with Et₂O, filtered through a pad of Plorisil, and the Florisil layer was washed with E_z . The combined filtrate and washings were concentrated in vacuo. The residue (15.8 g) was distilled in the presence of $x_2\omega_3$ to give 8 (10.8 g, 64 %), b.p. 92-94°/9 Torr; n_0^{24} 1.4438; vmax 3430 (s), 1130, (s), 1060 (s) cm^{-1} ; 6 (CCl₄) 1.18 (~2.4H, t, J=7 Hz), 1.25 (~0.6H, t, J=7 Hz,), 1.35~2.40 (4H, m), 2.87~4.30 (6H, m), 4.52 (~0.2H, dd, J=3, 14 Hz), 4.78 (~0.8H, dd, J=3, 3 Hz). (Found: C, 57.41; H, 9.55. Calc for C7H₁₄O₃: C, 57.51; H, 9.65 t).
2-Ethosytetrahydropyran-4-one 9. According to the reported procedure, 20.22 8 (4.7 g, 32 mmol) was oxidized with pyr

dichromate (PDC, 24.2 g, 64.3 mmol) in CH₂Cl₂ (90 ml) in the presence of MS 3A (34 g) followed by chromatographic purification [SiO₂ (80 g), n-hexane-Et₂O (10:1)] then distillation to give 9 (3.3 g, 72 %), h.p. 104~107°/23 Torr; n²⁵ 1.4395; vmax 1730 (s), 1120 (s), 1060 (s) cm⁻¹; 6 (CCl₄) 1.21 (3H, t, J=7 Hz), 1.90~2.83 (4H, m), 3.10~4.35 (4H, m), 5.05 (1H, dd, J=3.6, 3.6 Hz). (Found: C, 58.11; H, 8.38. Calc for C₇H₁₂O₃: C, 58.31; H, 8.39 %).

Ethyl 2-Ethoxy-4-oxo-5-tetrahydropyrancarboxylate 10 and ethyl 2-ethoxy-4-oxo-3-tetrahydropyrancarboxylate 11. A soln of LDA was prepared from n-BuLi (1.65 M in n-hexane, 12.4 ml, 20.5 mmol) and i-Pr₂MH (2.9 g, 20.5 mmol) in THF (45 ml) under Ar in a usual manner. To this soln was added dropwise a soln of 9 (25 g, 17.3 mmol) in THF (15 ml) over 13 min at -76° ~ 72° and then added HMPA (12 ml) over 2 min at -76°-66°. The reaction temp was allowed to rise to -35° to give a homogeneous soln. To this was added a soln of ethyl cyanoformate²⁴ (2,2 g, 21,8 mmol) in HMPA (2 ml) all at once at -76° \sim 56°. After stirring for 15 min at -78°, the mixture was poured into ice-water, neutralized by the addition of dil HCl and extracted with Et₂O. The extract was washed with brine, dried (MgSO₄) and concentrated in vacuo. The residue (9.1 g) was chromatographed over SiO₂ (110 g, n-hexane-EtOAc (17:1)) followed by the purification of fractions contaminated with 11 by prep TLC (C_GH_G-EtOAc (5.1)] to give 10 (total 2.04 g, 55 t). Analytical sample: b.p. 96-99°/0.45 Torr; n²⁰ 1.4656; vmax
1750 (s), 1730 (s), 1670 (s), 1635 (s), 1240 (s), 1060 (s) cm⁻¹; δ (CCl₄) 1.20 (3H, t, J each t, J=7 Hz), 1.90~2.80 (2H, m), 3.10~4.10 (2H, m), 4.17 (4H, q, J=7 Hz), 4.84 (1H, dd, J=2, 3 Hz), 5.12 (~0,27H, br.s), 11.77 (~0.53H, s), 12.12 (~0.2H, s). (Found: C, 55.18; H, 7.02. Calc for C₁₀H₁₆O₅: C, 55.54; H, 7.46 t). Further elution with the same solvent gave 11 (total 0.20 g, 5.4 %); vmax 1745 (s), 1680 (s), 1640 (w), 1565 (s), 1265 (s), 1150 (s) cm⁻¹; δ (CCl4) 1.25 (3H, t, J=7 Hz), 1.35 (3H, t, J=7 Hz), 1.90∼2.70 (2H, m), 3.20∼4.10 (3H, m), 4.02 (2H, q, J=7 Hz), 4.28 (2H, q, J=7 Hz), 5.07 (1H, br.d, J=5 Hz); MS m/z 216 (M⁺), 215, 201, 171, 143, 73, 45, 43 (base peak).

Ethyl (25,45,5R)- and (2R,4S,5R)-2-ethoxy-4-hydroxy-5-tetrahydropyrancarboxylate 12a. To a soln of sucrose (250 g, 730 mmol) in phosphate buffer (01 M, pH 8, 1.3 1) was added dry baker's yeast (110 g, Oriental Yeast Co., Ltd.) at 30°. After vigorous stirring for 10 min with aeration, to this fermenting mixture was added an emulsion of 10 (5,2 q, 24 mmol) in 0.2 \$ Triton X-100 ag soln (50 ml), and the stirring was continued for 1 h at 30°. After the addition of Et₂O, Celite and a small amount of Norit, the mixture was filtered. The filter-cake was washed thoroughly with EtOAc. The organic layer of the combined filtrate and washings was separated. The ag layer was saturated with NaCl and extracted with EtOAc. The combined EtOAc soln was dried (Mg9O₄) and concentrated in vacuo. The residue (7,8 g) was chromatographed over SiO₂ (80 g).
Firstly eluted fractions (n-hexane-EtOAc (15:1~12:1)) gave (25.45.5E)-12a (1.95 g, 37 %), n²⁰ $(\infty 0.51)$; vmax 3550 (s), 1730 (s), 1120 (s), 1100 (s), 1030 (s) cm^{-1} ; $\frac{1}{6}$ (100 MHz +D₂O) 1.26 (3H, t, J=7.2 Hz), 1.29 (3H, t, J=7.2 Hz), 1.87 (1H, ddd, J=3.4, 3.5, 14.2 Hz), 2.04 (1H, ddd, J=2.0, 3.7, 14.2 Hz), 2.72 (1H, ddd, J=2.9, 5.0, 12.2 Hz), 3.46 (2H, dq, J=7.2, 9.6 Hz), 3.76 (1H, dd, J=5.0, 11.7 Hz), 3.67~3.98 (1H, m), 4.18 (1H, dd, J=11.7, 12.2 Hz), 4.20 (2H, q, J=7.2 Hz), 4.20 (2H, dq, J=1.7, 12.2 Hz), 4.20 (2H, the optical purity of (25,45,58)-12a was as follows. (a) According to the reported procedure²⁷, (25,45,58)-12a was converted to its (R)- and (S)-MTPA ester, (2S,4S,5R)-12b and 12c, respostively. (b) HPLC analysis of (2S,4S,5R)-12b: [Column, Silica-1251N, 25 cm x 4.6 mm; n-hexane-THF (20:1), 1 ml/min) Rt 56.6 min (28.5 %), 57.9 min (71.5 %), HFLC analysis of the \sim 1:1 mixture of (2S,4S,5R)-12b and 12c under the same condition as above showed Rt 59,7 min (48,4 %) and 60,0 min (51,6 %). Therefore the optical purity of (2S,4S,5R)-12a was determined to be 43 % e.e.

Secondly eluted fractions (n-hexane-stok: (10:1-6:1)) gave a mixture of (25,45,5R)- and (2R,45,5R)-12a (1.60 g, 31 %). Thirdly eluted fractions $\frac{1}{2}$ = because Etokc (8:1)] gave (2R,4S,5R)-12a (0.60 g, 11 s), m_1^3 1.4549; $\frac{1}{2}$ (a) $\frac{1}{2}$ -76.9° (c=0.54); vmax 3500 (s), 1735 (s), 1140 (s), 1065 (s) cm⁻¹; δ (CCl₄) 1.16 (3H, t, J-7 Hz), 1.28 (3H, t, J-7 Hz), 1.55-2.00 (2H, m), 2.62 (1H, ddd, J=3.2, 4.0, 4.4 Hz), 3.03 (1H, br.d, J=5.8 Hz, OH), 3.20~4.10 (5H, m), 4.16 (2H, q, J=7 Hz), 4.71 (1H, dd, J=3.6, 4.4 Hz); HR-MS m/2 217.1087 (M⁺-H), Calc for C₁₀H₁₇0₅: 217.1076, Since the separation of the diastereomeric mixture of (28,45,58)-12b and 12c by HFLC was difficult, the optical purity of (28,45,58)-12a was determined as follows. (a) Preparation of an equilibrium mixture of (2RS,4S,5R)-12a. A soln of (2R,4S,5R)-12a (9.5 mg, 0.043 mmol) and a catalytic amount of p-TSOH.H₂O in 99 & EtOH (0,5 ml) was stirred for a week at room temp and then diluted with Et₂O, washed with sat NaHCO₃ ag soln and brine, dried (MgSO₄) and concentrated in vacuo to give (2RS₄S₂,5R)-12a (10.9 mg). (b) HPLC analysis of
the MTPA ester, (2RS₂,4S₂,5R)-12b, prepared from (2RS₂,4S₂,5R)-12a and (S)-MTPA-Cl for that of (2S,4S,5R)-12b) Rt 37.3 min (70.8 %, (2R,4S,5R)-12b), 58.0 min (28.3 %, (2S,4S,5R)-12b). (2S,4S,5R)-12b was shown to be a single diastereomer. Therefore the optical purity of (2R,4S,5R)-12a was determined to be 100 % e.e.

Ethyl (45,5R)-4-Acetoxy-2-ethoxy-5-tetrahydropyrancarboxylate 12d. (2RS,4S,5R)-12a (4.7 g, 22 mmol) was treated with Ac₂O (6.2 ml, 65.7 mmol) and 4-(N,N-dimethylamino)pyridine (DMAP, 0.25 g, 2.0 mmol) in C₅H_cN (20 ml) in a usual manner followed by chromatographic purification over SiO₂ (70 g, m-hexane-EtOAc (9:1)) to give 12d (6.0 g, quantitative). A small amount of $(25.45.55)$ -12d was obtained from the later eluted fractions: n_0^{16} 1.4419; [a] n_0^{17} +32.7° (c=1.07); vmax 1740 (s), 1240 (s), 1125 (s), 1025 (s) cm⁻¹; 6 (ccl₄) 1.19 (3H, t, J-7 Hz), 1.23 (3H, t, J-7 Hz), 1.97 (3H, s), 1.70-2.20 (1H, m), 2.20-2.49 (1H, m), 2.66 (1H, ddd, J=3.2, 4.8, 11 Hz), 3.00~4.20 (4H, m), 4.12 (2H, q, J=7 Hz), 4.67 (1H, dd, J=2, 4 Hz), 5.21 (1H, ddd, J=3.2, 3.2, 3.2 Hz). (Found: C, 55.30; H, 7.73. Calc for C₁₂H₂₀O₆: C, 55.37; H, 7.75 %).

(45,5R)-2-Ethoxy-4-hydroxy-5-tetrahydropyranmethanol 13a. 12d (5.8 g, 22.3 mmol) was reduced with LAH (1.75 g, 46.1 mmol) in Et₂0 (100 ml) in a usual manner to give crude 13a (3.8 g, 97 %), wmax 3420 (s), 1120 (s), 1030 (s) cm⁻¹; 6 (CCl₄) 1.25 (3H, t, J=7 Hz), 1.45 (2H, s, OHx2), 1.55~2.40 (3H, m), 2.80~4.60 (7H, m), 4.95 (1H, br.dd, J=3, 3 Hz); MS m/z 176 (M⁺). This was employed in the next step without further purification.

(2R, 4S, 5S) - and (2S, 4S, 5S) - 2-ethoxy-4-hydroxy-5-tetrahydrogyranmethanol dibenzoate 13c. 13a (44 mg, 0.25 mmol) was treated with benzoyl chloride (0.12 ml, d 1.211, 1.0 mmol) in C5H5N (0.7 ml) in a usual manner followed by chromatographic purification over SiO₂ [2 g, n-hexane-EtOAc (40:1~30:1)] to give (2RS, 4S, 5S)-13c (91 mg, 95 %). HPLC (n-hexane-THF (20:1), 1.1 ml/min] Rt 31.9 min (37 %), 52.7 min (63 %). Further purification of this by prep TLC [n-hexane-Et₂O (5:1, x 8)] gave $(2E, 4S, 5S)$ -13c (19 mg, 21 %) as a less polar isomer and $(2S, 4S, 5S)$ -13c (47 mg, 52 %) as a more polar isomer. $(2E, 4S, 5S)$ -
13c; TLC [n-hexane-EtOAc (2:1)] Rf 0.66; CD (-9.0 x 10⁻³, EtOH, t=25°): [ΔE (λ, nm)] + 3090 (w), 3040 (w), 1725 (vs), 1605 (m), 1590 (m), 1490 (m), 1270 (vs), 1110 (vs), 715 (vs), 690 (m) cm⁻¹, 6 (100 MHz) 1.26 (3H, t, J=7.3 Hz), 1.97 (1H, ddd, J=4.4, 4.4, 13 Hz), 2.17 (1H, ddd, J=3.3, 7.3, 13.0), 2.40~2.77 (1H, m), 3.50 (1H, dq, J=7.3, 10 Hz), 3.84 (1H, dq, J=10, 7.3 Hz), 3.87 (1H, dd, J=5.8, 12 Hz), 4.11 (1H, dd, J=4.4, 12 Hz), 4.52 (2H, d, J=7.3 Hz), 4.91 (1H, dd, J=3.3, 4.4 Hz), 5.70 (1H, ddd, J=4.1, 4.4, 7.3 Hz), 7.20~7.70 (6H, m), 8.02 (4H, ddd, J=2.0, 7.3, 9.3
Hz), A.91 (1H, dd, J=3.3, 4.4 Hz), 5.70 (1H, ddd, J=4.1, 4.4, 7.3 Hz), 7.20~7.70 (6H, m), 8.02 (4H, 1585 (m), 1495 (w), 1270 (vs), 1110 (vs), 710 (s), 690 (m) cm⁻¹, 6 (100 MHz) 1.21 (3H, t, J=7.3 Hz), 2.04 (1H, ddd, J=4.9, 5.8, 13 Hz), 2.17 (1H, ddd, J=4.9, 5.8, 13 Hz), 2.45~2.75 (1H, m), 3.45 (1H, dq, J=10, 7.3 Hz), 3.75 (1H, dd, J=8.7, 12 Hz), 3.86 (1H, dq, J=10, 7.3 Hz), 4.24 (1H, dd, J=8.4, 12 Hz), 4.43 (2H, d, J=7.3 Hz), 4.78 (1H, dd, J=4.9, 4.9 Hz), 5.47 (1H, ddd, J-5.8, 5.8, 5.8 Hz), 7.12-7.70 (6H, m), 8.04 (4H, ddd, J-2.0, 7.3, 9.3 Hz); MS m/z 384 (M⁺), 369, 339, 262, 217, 140, 123, 122, 105, 77.

(45,5R)-2-Ethoxy-4-hydroxy-5-tetrahydropyranmethanol dibenzyl ether 13b. Under Ar 13a (3.7 g, 21 mmol) was treated with NaH (3.5 g, 60 % in mineral oil, 87.5 mmol), n-Bu_dN⁺I (0.38 g, 1.0 mmol)³⁰ and PhCH₂Br (10.9 g, 63.8 mmol) in THF (80 ml) with refluxing for 1 h followed by chromatographic purification over SiO₂ [70 g, n-hexane-BtOAc (50:1-10:1)] to give 13c
(6.0 g, 80 t), n₀⁷ 1.5267; [a]₀²¹ -16.3° (c=0.71); vmax 3110 (w), 3080 (m), 3050 (m), 1500 cm⁻¹; 6 (CCl₄) 1.15 (3H, t, J=7 Hz), 1.50~1.87 (2H, m), 1.87~2.33 (1H, m), 3.00~4.21 (7H, m), 4.39 (4H, br.s), 4.49~4.82 (1H, m), 7,17 (10H, br.s); MS m/z 356 (M⁺), 341, 310, 241, 219, 203, 202, 91.

 $(45,5R)-4-Benzyloxy-5-benzyloxymethyl-2-tetrahydroyzanyltriplemylospionium tetrafluoroborate 14. A soln of 13b (1,13 q, 3,17 mmol) and Ph₃FH⁺BF₄ (1,26 q, 3,61 mmol)³¹ in MeCN (20 ml) was stirred and heated under reflux for 1 h under Ar. The$ mixture was concentrated in vacuo, and the residue was washed with Et₂0 (10 ml x 13). The residue was then suspended in C₆H₆ (20 ml), and the mixture was concentrated in vacuo to remove H₂O azeotropically. This process was repeated four times. The residue was heated at ~00° (bath temp) in vacuo overnight to dryness, giving crude 14 (2.20 g, quantitative). This was employed in the next step without further purification.

Ethyl (25,1'5)-2-(1'-hydroxyethyl)pent-4-enoate (25,1'5)-17a According to the reported procedure, 33 (5)-16 [b.p. 70-75°/15 Torr; $\left[\frac{1}{6}\right]^{2}$ +42.6° (c=1.04); 96-98 **0** e.e.; 26.4 g, 0.20 moll gave (25.1'S)-17a (29.3 g, 85.2 **a**), b.p. 95-103°/16 Torr; $\left[\frac{a}{b}\right]^{2}$ +14.8° (c=1.55); $\left[11t^{33}$ $\left[\frac{a}{b}\right]^{2}$ +14.5° (cm0.37, GE1₃)

Ethyl (2RS, 1RS)-2-(1'-hydroxyethyl)pent-4-enoate (2RS, 1RS)-17a. Ethyl 2-acetylpent-4-enoate [5.4 g, prepared from ethyl acetoacetate 15, allyl bromide and K₂O₃ in refluxing acetone-DMF (4:1); contaminated with dialkylated product (purity 60 %) was reduced with NaBH₄ (0.49 g, 10.3 mmol) in 99 % EtOH (30 ml) in a usual manner. Chromatographic purification (SiO₂) (140 g), n-hexane-Et₂O (12:1~10:1)] gave ethyl 2-allyl-2-(1'-hydroxyethyl)pent-4-enoate (21 g, 26 % from 15) and (t)-17a $(2.1 g, h.p. 61-65^{\circ}/3 Torr; 32$ \$ from 15).

Ethyl (2S,1'S)-2-(1'-tetrahydropyranyloxyethyl)pent-4-enoate 17b. (2S,1'S)-17a (29,1 g, 169 mmol) was treated with dihydropyzan (23,1 g, 274 mmol) and pyridinium p-toluenesulfonate (PPTS, 5.5 g, 21.9 mmol) in CH₂Cl₂ (150 ml) followed by a usual workup^{Cf.10},¹¹ to give 17b (43,7 g, quantitative), b.p. 92°/0.2 Torr; no²⁴ 1.4476; [a (m), 1740 (s), 1650 (m), 1185 (s), 1125 (s), 1035 (s), 1025 (s), 990 (s), 915 (s) cm⁻¹; 6 (CCl₄) 1.11 (3H, d, J=7 Hz), 1.24 (3H, t, J=7 Hz), 1.35~2.00 (6H, m), 2,36 (2H, br.d, J=5 Hz), 1.97~2.65 (1H, m), 3.10~4.40 (3H, m), 4.08 and 4.10 (total 2H, each q, J=7 Hz), 4.45~4.78 (1H, br.d, J=8 Hz), 4.78~5.25 (2H, m), 5.25~6.30 (1H, m). (Found: C, 65.29; H, 9.44. Calc for $C_{14}H_{24}O_4$: C, 65.69; H, 9.44 %).

 $\frac{(2R,35)-2-\text{Allylbutane-l,3-diol} 3-\text{MIP} \text{ either}}{450 \text{ ml}}$ 17b (43.5 g, 169.7 mmol) was reduced with LAH (7.5 g, 198 mmol) in Et₂O (2R, 3S)-2-Allylbutane-1, 3-diol 3-TMP ether 18a. 17b (43.5 g, 169.7 mmol) was reduced with LAH (7 vmax 3470 (s), 3100 (m), 1645 (m), 1025 (s), 995 (s), 910 (m) cm⁻¹; 6 (CCl₄) 1.15 and 1.25 (total 3H, each d, J=7 Hz), 1.25-1.95 (7H, m), 2.12 (2H, dd, J-7, 7 Hz), 2.45 and 2.57 (total 1H, each dd, J-5, 5; 5, 7 Hz, OH), 3.12-4.30 (5H, m), 4.40~4.75 (1H, br.s), 4.75~5.35 (2H, m), 5.35~6.30 (1H, m). (Found: C, 66.84; H, 10.56. Calc for C₁₂H₂₂O3: C, 67.25; H,

 10.35 a).

(<u>2R,3S)-2-Allylbutane-1,3-diol 1-benzyl 3-THP ether</u> **182. 18a** (36.2 g, 169 mmol) was treated with NaH (1L8 g, 60 % in
mineral oil, 295 mmol), <u>n</u>-Bu₄N⁺I⁻ (1.15 g, 3.1 mmol)³⁰ and PhCH₂Br (39.5 g, 231 mmol) in h under Ar followed by a usual workup to give crude 18b (59.7 g). A portion (3 g) was chromatographed over SiO₂ [6Og, nhexane-BtOAc (100:1~70:1~50:1)] followed by distillation in the presence of K₂OO₃ to give an analytical sample, b.p. $137^{\circ}/0.3$ Torr; n_0^2 1.4986; $\left[\alpha\right]_0^2$ +12.8° (c=1.01); vmax 3100 (w), 3060 (w), 1645 (m), 1505 (m), 1120 (a), 1080 (s), 1025 (s), 995 (a), 910 (m), 740 (m), 705 (s) cm⁻¹; $\frac{1}{2}$ (CC1₄) 1.05 and 1.15 (total 3H, each d, J=7 Hz), 1.25-1.75 (6H, br.m), 1.65-2.05 (lli, ml, 2.11 (ZH, br.dd, J-6, 7 Hz), 3.37 and 3.44 (total 2H. each d, J-5 Hz), 3.55-4.05 (2H. m), 3.87 (IH, m), 4.41 (ZH, s), 4.45-4.73 (1H. br), 4.7--5.2 (ZH, m), 5.3-6.2 (Xi, m), 7.28 (5H. br.s). Wound: c, 74.901 H, 9.19. ca1c for $C_19H_28O_3$: C, 74.96; H, 9.27 a).

(2R_t3S)-2-Allylbutane-1,3-diol 1-benzyl ether 18c. (2R₅3S)-18b (59.0 g, ~167 mmol) was treated with p-TsOH·H₂O (L15 g, 6.0 mmol) in MeOH (500 ml). After a usual workup,^{CI.11} chromatographic purification [SiO₂ (700 g), <u>n</u>-hexane-EtOAc (10:1-8:1)1 gave **l&z** (38.4 g, quantitative). AMlytical sample: hp 108'/O.l7 Parr, d4 150568 **l.li\$3** -1.50' (c=l.47)1 vmax 3520 (s), 3100 (m), 3060 (m), 3010 (s), 1645 (m), 1500 (m), 1010 (s), 1000 (s), 915 (s), 740 (s), 700 (s) cm⁻¹; 6 $(CC1₄)$ 1.12 (3H, d, J-7 Hz), 1.36-1.85 (1H, m), 2.12 (2H, br.dd, J-6, 7 Hz), 2.55-2.75 (1H, m, OH), 3.46 (2H, dd, J-3, 5 Hz), 3.71 (1H. dq, J=6, 6 Hz), 4.41 (2H. s), 4.72-5.20 (2H, ml, 5.35-6.15 (1H. m), 7.27 (5H. br.ss)t Capillary GLC (Column. PBG 2OM, 50 m x 0.25 mm at 170°; Carrier gas, N₂, 1.1 kg/cm²) Rt 38.8 min (96.7 %), 39.9 min (3.3 %). (Pound: C, 75.96; H, 9.01. Calc for $C_{14}H_{20}O_2$: C, 76.32; H, 9.15 0).

(2R₂3S)-2-Allylbutane-1,3-diol 1-benzyl ether 3-(3',5'-dinitro)benzoate 10d. 16c (36.9 g, ~159 mmol) was treated with 3,5dinitrobenzovl chloride (DNB-Cl, 40.4 g, 175 mmol) and DMAP (0.65 g, 5.3 mmol) in C₅H_eN (140 ml) in a usual manner^{cf.11} to give crude 18d (60.7 g, 92 %). This was recrystallized four times from n-hexane-Et₂O (3:1) followed by another recrystallization from n-hexane-Et₂O (100:1) to give pure 18d (44.6 g, 73.5 % recovery) as yellowish needles, m.p. 61-62°, [a]²⁴ +42.1' (c=2.50), umax 3150 (~1, 3120 (ml, 1720 (81, 1655 (~1, 1630 cm), 1600 (w), 1550 (81, 1500 (~1, 1350(s), 1280 (9). 1180 (a), 925 (s), 880 (m), 760 (m), 735 (s), 730 (s), 720 (s), 705 (m) cm⁻¹; δ (CDC13) 1.44 (3H, d, J=6 Hz), 1.90~2.50 (1H. m), 2.20 (2H. dd, 516, 6 Hz), 3.55 (ZH, brd, J-5 Hz), 4.45 (ZH, 81, 4.86-5.30 (ZH, IO), 5.30-6.20 (2H, ml, 5.40-6.25 (lH, ml, 5.55 (IH, dq, J=6, 6 Hz), 7.25 (5H. br.s), 9.10 (ZH, d, J-2 Hz), 9.21 (1H. t, J-2 Hz). HPLC ["-hexane-THF (1OO:l). 1.3 ml/mini Rt 44.2 min (99.9 a). Its diastereomer eluted at Rt 42.3 min. (Pound: C, 61.17; **H, 5.39; N.** 6.75. Calc for $C_{21}H_{22}O_7N_2$: C, 60.96; H, 5.35; N, 6.76 **8**).

Diastereomerically and enantiomerically pure $(2R,3S)-2-\lambda 11$ yltutane-1,3-diol 1-benzyl ether 18c. $(2R,3S)-18d$ $(44.6 g, 108)$ mmol) was hydrolyzed with N KOH aq soln (150 ml. 150 mmol) in THF-99 \$ StOH (1:1, 360 ml)^{cf-11} to give 18c (23.5 g, 99 %), b.p. 108~109°/0.16 Torr; m_0^2 ⁶ 1.5045; [a] $\frac{2}{5}$ -1.96° (c=2.08); its IR and NMR spectra were almost identical with those deacribed above. Capillary GLC (under the same condition as described above) Rt 18.4 min (100 %). Its diastereomer eluted at Rt 19.7 min. (Found: C, 76.28; H, 9.17. Calc for $C_{14}H_{20}O_2$: C, 76.32; H, 9.15 W. According to the reported procedure.²⁷ (2R,3S)-18e was prepared from (2R,3S)-18c and (S)-HTPA chloride (MTPA-Cl). HPLC [n-hexane-THF (100:1), 1.3 ml/min] Rt 18.0 min [single peak, (2R,3S)-18e]. Its diastereomer (2S,3R)-18e prepared from (2RS,1%S)-17a eluted at Rt 19.2 min.

(2R,3S)-2-Allylbutane-1,3-diol 1-benzyl ether 3-tosylate **18f. (2R,3S)-18c (12 g, 54.5 mmol) was** treated with p-TsCl (25 g, 130 mmol) and DMAP (1.46 g, 12 mm011 in CgHgN 00 ml) in a &l-&"&fJo*ll to give **18f** (20.5 g, quantitative). ?his was employed in the next step without further purification; vmax 3100 (w), 3060 (w), 3020 (m), 1645 (m), 1605 (m), 1500 (w), 1360 (m), 1180 (s), 1100 (s), 910 (s), 815 (m), 780 (m), 735 (m), 700 (m) cm⁻¹; δ (CCl₄) 1.21 (3H, d, J=7 Hz), 1.61~2.28 (3H, m), 2.38 (3H, s), 3.29 (2H, d, J=7 Hz), 4.30 (2H, s), 4.50~5.20 (3H, m), 5.25~6.10 (1H, m), 7.25 (2H, d, J=8 Hz), 7.26 (5H, br.s), 7.75 (2H, d, J=8 Hz).

(R)-5-Benzyloxy-4-ethyl-l-pentene 19a. Super Hydride[®] (LiBEt₃H, 1 M in THF, 64 ml, 64 mmol) was added dropwise to a stirred soln of 18f (121 g, 32 mmol) in THF (36 ml) below -5^o under Ar. The mixture was stirred and heated under reflux for 20 min. After cooling, to the mixture were added H₂O (5 ml), 3 N NaOH aq soln (30 ml) and 35 * H₂O₂ aq soln (27 ml) successively below 50° with 10e-cooling, The mixture was stirred for 30 min at room temp, then concentrated in vacuo to remove THF and EtOH. The residue was extracted with Et₂0 (50 ml x 5). The extract was washed with H₂O and brine, dried (MgSO₄) and concentrated <u>in vacuo</u>. The residue (7.0 g) was chromatographed over SiO₂ (70 g n-pentane-Et₂O (20:1)] to give
19a (6.5 g, quantitative). Analytical sample: b.p. 90°/0.3 Torr; n²⁵ 1.4892; [α]24 +4.62° (w), 3010 (w), 3000 (s), 1645 (m), 1500 (w), 1100 (s), 995 (m), 910 (s), 735 (s), 700 (s) cm⁻¹; 6 (CC1₄) 0.86 (3H, t, J-6 Hz), 1.08-1.85 (3H. m), 2.11 (ZH, dd, J-6, 6 Hz), 3.30 (ZH, d, J-5 Hz), 4.41 (2H. s), 4.75-5.22 (ZH, ml, 5.40-6.20 (lH, ml, 7.30 (5H, br.s). (Found: C, 82.34; H, 9.70. Calc for $C_{14}H_{20}O$: C, 82.30; H, 9.87 t).

(RI-2-Ethyl-4-penten-l-01 **(lJ)-19& lb a sol" of @J-190 6.5 g,** 32 mmol) in ?w (16 ml) arxl liq. NH3 t-63 ml) was added finely cut Na (1.66 g, 72.2 mmol) portionwise with dry ice-acetone cooling and stirring. The mixture was refluxed for 10 min with stirring. To the dry ice-acetone cooled muxture was added solid NH4Cl to discharge its deep blue color. NH₃ was evaporated from the mixture with stirring at room temp. The residue was diluted with Et₂O and filtered through a pad of Celite. The solid was washed with Et₂O. The combined filtrate and washings were washed with brine, dried (Mg90₄) and concentrated <u>in vacuo</u> to give crude (<u>R</u>)-19b (7.3 g, contaminated with PhMe). Analytical sample: b.p. 92~93°/53 Torr; nß° 1.4394; [a] $_0^1$ ³ -0.75° (c=1.56); vmax 3350 (s), 3070 (m), 1655 (s), 1040 (s), 990 (s), 910 (s) cm⁻¹; δ (CC1₄) 0.88 (3H, t, J-6 Hz), 1.05-1.60 (3H. m), 2.06 (ZH, dd, J-6, 6 Hz), 2.86 (lH, br.t, 515 Hz, OH), 2.43 (2H. br.dd, J=5, 5 Hz), 4.65-5.25 (ZH, ml, 5.30-6.20 (1H.m). (Found: C, 73.441 **H.** 12.21. Calc for **C7H140: C,** 73.64, **H,** 12.36 t).

(R)-4-Ethyl-5-tetrahydropyranyloxy-1-pentene 19c. Crude 19b (7.3 g, -32 mmol) was treated with dihydropyran (9.0 g, 107 mmol) and PPTS (4.0 g, 16 mmol) in CH₂Cl₂ (45 ml) as described above followed by chromatographic purification [SiO₂ (70 g), n -pentane-Et₂O (3O:1)) then distillation in the presence of K₂CO₃ to give 19c (6.1 g, quantitative from pure (2R,3S)-18cl, \bar{h} p 77°/3 Torr; n_0^2 1.4481; [a] β ¹ +3.00° (c=1.07); vmax 3080 (m), 1640 (m), 1120 (s), 1030 (s), 990 (s), 910 (s) cm⁻¹; 6 $(CCl₄)$ 0.92 (3H, t, J=6 Hz), 1.15~2.00 (9H, m), 2.12 (2H, dd, J=6, 6 Hz), 2.98~4.10 (4H, m), 4.40~4.65 (1H, br.s), 4.75~5.30 (2H, m), 5.40~6.20 (1H, m). (Found: C, 72.62; H, 11.14. Calc for C₁₂H₂₂O₂: C, 72.62; H, 11.18 %).

(R)-3-(Tetrahydropyranyloxymethyl)pentanal 20. To a strred two-phase mixture of 19c (1.5 g, 7.9 mmol) in Et₂O-H₂O (1.1, 40 InI) were xkled MI04 (5.6 g, 26.2 mmol) and 5 \ Cm04 soln in 'IWF (2.4 ml, **0d7** mwl) at room temp Aftar stirrirsg vigorously for 2 h at room temp, an additional amount of NaIO₄ (3 g, 14 mmol) was added to the mixture. The vigorous strrring was further continued for 5.5 h at room temp. The precipitate was filtered off through a pad of Celite and washed with Et₂O. The organic layer was separated from the combined filtrate and washings, and the aq layer was extracted with Bt₂0. The combined Et₂O soln was washed with Na₂S₂O₃ aq soln, sat NaHOO₃ aq soln and brine, dried (MgSO₄) and concentrated in vacuo. The residue was distilled under Ar to give 20 (1.02 g, 65 a), b.p. 94°/2.0 Torr; n_0^2 1.4529; $\left[\frac{a}{2} + 25.2^{\circ}\right]$ $\overline{(c=1.07)}$; vmax 2740 (m), 1725 (s), 1125 (s), 1035 (s) \overline{c} cm⁻¹; 6 (ccl₄) 0.92 (3H, t, J=7 Hz), 1.15~2.15 (9H, m), 2.15~2.50 (2H. ml, 2.80-3.96 (QH, m), 4.30-4.60 (lH, br.6). 9.74 and 9.79 (total 1H. each d, J-2 Hz). (Found: C, 66.078 H, 10.08. Calc for $C_{11}H_{20}O_{3}$: C, 65.97; H, 10.07 8).

(3'R,4S,5R)-4-Benzyloxy-5-benzyloxymethyl-2-((3'-tetrahydropyranyloxymethyl)pentylidene]tetrahydropyran 22. To a stirred and cooled soln of crude 14 (2.20 g, ~ 3.17 mmol) and HMPA (5.5 ml) in THF (29 ml) was added dropwise n-Buli (1.72 M in nhexane, 21 ml, 3.61 mmol) at -67°-63° under Ar. After stirring for 30 min at -67°, to this deep red ylide soln was added dropwise a soln of 20 (0.83 q, 4.12 mmol) in THF (6.5 ml) at $-67^{\circ} - 63^{\circ}$ with stirring. The temp was raised gradually to room temp over 3 h 50 min and the stirring was continued for 6 h 15 min at room temp. To the mixture was added sat NaHCO3 ag soln (18 ml) at -10°~0° with stirring. The mixture was concentrated in vacuo to remove THF. The residue was extracted with Et₂O (20 ml x 4). The extract was washed with sat NaHOO3 aq soln and brine, dried (K₂OO3) and concentrated in vacuo to give crude 22 (2.94 g). According to the procedure described above, 3.85 g (10.8 mmol) of 13b was converted to 9.0 g of crude 22. This was employed in the next step without further purification.

(3R,4S,6R,9R)- and (3S,4R,6S,9R)-4-Benzyloxy-3-benzyloxymethyl-9-ethyl-1,7-dioxaspiro[5.5]undecane 1b + 23b and (3R, 4S, 6R, 9R)-, (3S, 4S, 6R, 9R)- and (3R, 4R, 6S, 9R)-3-benzyloxymethyl-9-ethyl-4-hydroxy-1,7-dioxaspiro(5.5)undecane 1c + 2c + 24c. Crude 22 (9.0 g) was mixed with conc HCl-H₂O-THF (1:5:20, 90 ml) with ice-salt cooling. After stirring for 15 h at room temp, the mixture was neutralized by the addition of sat NaHOO3 aq soln with stirring and ice-cooling, and concentrated in vacuo to remove THF. The residue was extracted with Et₂0. The extract was washed with sat NaHCO₃ aq soln and brine, dried (MgSO₄) and concentrated in vacuo. The residue (7.8 g) was chromatographed over SiO₂ [210 g, n-hexane-EtOAc (80:1-10:1)] to give a mixture (2.04 g, 46 \from 13b) of 1b and 23b, n_1^{24} 1.5274; $\left(\frac{n}{2}\right)^{2}$ -105° (c=0.51); vmax 3100 (w), 3080 (w), 3050 (m), 1500 (m), 1105 (vs), 1080 (vs), 1005 (s), 740 (s), 700 (s) cm⁻¹; 6 (100 MHz) 0.88 and 0.90 (total 3H, each t, J=7.5 Hz), 1.00-1.80 (8H, m), 1.87 (1H, dd, J=5.2, 13.3 Hz), 2.10~2.43 (1H, m), 3.14 (~0.7H, dd, J=9.9, 9.9 Hz), 3.27~3.63 (~2.3H, m), 3.73 (2H, d, J=6.9 Hz), 4.95 and 4.97 (total 1H, each dd, J=3.7, 11.7; 4.0, 12.0 Hz), 4.06 (1H, ddd, J=5.1, 5.1, 11.0 Hz), 4.43 (1H, d, J=11.7 Hz), 4.47 (1H, d, J=11.7 Hz), 4.59 (1H, d, J=11 Hz), 4.62 (1H, d, J=11.7 Hz), 7.28 (~6H, br.s), 7.33 (~4H, br.s). (Found: C, 75.63; H, 8.38. Calc for C₂₆H₃₄O₄: C, 76.06; H, 8.34 %). Further elution [n-hexane-EtOAc (10:1~3:1)] gave a mixture (1.21 g, 35 % from 13b) of 1c, 2c and 24c, vmax 3430 (s), 3100 (w), 3070 (w), 3040 (w), 1500 (w), 1075 (s), 1045 (s), 740 (m), 700 (m) cm⁻¹; δ (CCl_A) 0.88 (3H, t, J=6.4 Hz), 1.05~1.50 (10H, m), 2,90~4.30 (8H, m), 4.49 (2H, br.s), 7.32 (5H, br.s); MS m/z 320 (M⁺), 302, 129, 127, 126, 125, 91 (base peak).

(3R,4S,6R,9R)-, (3S,4S,6R,9R)- and (3R,4R,6S,9R)-4-[3-Benzyloxymethyl-9-ethyl-1,7-dioxaspiro(5.5 lundecyl) benzoate 1f, 2f and 24f. A mixture of 1c, 2c and 24c (25 mg, 0.078 mmol) was treated with benzoyl chloride (0.012 ml, d 1.211, 0.10 mmol) in C5H5N (0.5 ml) in a usual manner followed by purification with prep TLC (n-hexane-EtOAc (7:2)) to afford 28 mg (82 %) of 1f, 2f and 24f, vmax 3110 (w), 3080 (w), 3050 (w), 1725 (s), 1605 (w), 1590 (w), 1495 (w), 1270 (s), 1100 (s), 740 (m), 715 (s) cm⁻¹; 6 (100 MHz) 0.89, 0.91 and 0.96 (total 3H, each t, J=7.3 Hz), 1.00~2.00 (7H, m), 1.97 (1H, dd, J=5.1, 13.1 Hz), 2.10~2.53 (1H, m), 3.41 (~0.33H, dd, J=9.5, 9.5 Hz, 1f), 3.52 (~0.33H, dd, J=13.1, 13.1 Hz, 2f), 3.56 (~0.33H, dd, J=3.7, 13.1 Hz, 2f), 3.61 (~0.33H, br.d, J=11.7 Hz, 24f), 3.55~3.65 (~0.33H, m, 1f), 3.60~3.80 (2H, m), 3.75 (~0.33H, dd, J=5.1, 11.7 Hz, 24f), 3.73~3.82 (~0.66H, m, 1f), 3.87 (~0.67H, dd, J=11.7, 11.7 Hz, 2f + 24f), 3.96 (~0.67H, dd, J=5.1, 11.7 Hz, $2f + 24f$, 4.42 (~0.67H, s), 4.45 (~0.67H, s), 4.53 (~0.66H, s), 5.36 (~0.67H, ddd, J=5.1, 11, 11 Hz, $2f + 24f$), 5.62 (~0.33H, ddd, J=5.5, 5.5, 11.7 Hz, 1f), 7.26 and 7.29 (total 5H, each s), 7.36~7.75 (3H, m), 7.83~8.20 (2H, m), MS m/z 424 (M⁺), 423, 321, 312, 302, 129, 127, 126, 123, 122, 105, 91, 77.

(3R, 4S, 6R, 9R) - and (3S, 4R, 6S, 9R)-9-Ethyl-4-hydroxy-3-hydroxymethyl-1, 7-dioxaspiro(5.5) undecane la and 23a. A mixture of 1b and 23b (2.35 g, 5.72 mmol) was treated with Na (1.41 g, 61.3 mmol) in THF (6 ml) and liq. NH₃ (~80 ml) in the usual manner to give crude la and 23a (0,99 g, 75 %), wmax 3370 (s), 1070 (s) cm⁻¹; 6 (CDCl₃) 0,88 (3H, distorted t, J=7 Hz), 2.35 (2H, s, OHx2), 1,1-2.4 (10H, m), 2.73-4.10 (6H, m), 4.10-4.65 (1H, m). This was employed in the next step without further purification

(38,4S,6R,9R)-, (3R,4R,6S,9R)-, (3R,4R,6R,9R)- and (3S,4S,6S,9R)-3-[9-Ethyl-4-hydroxy-1,7-dioxaspiro[5.5]undecyl]methyl (3'5'-dinitro)benzoate le (talaromycin À 12-(3',5'-dinitro)benzoate), 23e, 25e and 26e. A mixture of crude la and 23a (0.99 g, 4.3 mmol) was treated with DNB-Cl (1.10 g, 4.75 mmol) and DMAP (16 mg, 0.13 mmol) in C₅H₆N (18 ml) with ice-salt cooling. The stirring was continued for 3 days at room temp. After a usual workup,¹¹ the residue (1.9 g) was chromatographed over SiO₂ (40 g). Firstly eluted fractions [C₆H₆-EtOAc (80:1-60:1)] gave 487 mg (18 %) of 1d and 23d as crystals, TLC [n-hexane-EtOAc (3:1)] Rf 0.53; vmax 3120 (m), 1730 (s), 1630 (m), 1600 (w), 1550 (s), 1350 (s), 1280 (s), 1165 (s), 760 (s), 730 (s), 725 (s) cm⁻¹; 6 (CDC1₃) 0.70~1.70 (3H, m), 1.10~2.15 (8H, m), 1.93~2.35 (1H, m), 2.35~2.90 (1H, m), 3.25~4.70 (4H, m), 4,77 (2H, br.d, J=6 Hz), 5,50-6,10 (1H, m), 9,00-9.37 (6H, m); MS m/2 406 (M⁺-DNBOH).

Secondly eluted fractions (40:1~10:1) gave 25e and 26e (254 mg, 14 %), TLC (n-hexane-EtOAc (3:1)) Rf 0.36; umax 3530 (s), 3120 (a), 1735 (s), 1630 (m), 1600 (w), 1550 (s), 1350 (s), 1280 (s), 1080 (s), 775 (m), 760 (m), 730 (s), 725 (s) cm⁻¹; 6 (CDCl₃) 0.89 (3H, br.t, J²⁵ Hz), 1.10~2.20 (8H, m), 2.00~2.70 (2H, m), 3.10~4.37 (5H, m), 4.44 and 4.50 (total 2H, each br.d. J=7.0 Hz), 4.65~5.10 (1H, m), 9.23 (2H, d, J=2 Hz), 9.30 (1H, t, J=2 Hz); MS m/z 424 (M⁺). 36 mg of this was further purified by prep TLC [n-hexane-EtOAc (3:1, x 2)] to afford 26e (16 mg, 44 %) as a less polar isomer and 25e (5 mg, 14 %) as a more polar isomer. 26e: TLC [n-hexane-EtOAc (3:1 x 3)] Rf 0.62; vmax 3530 (s), 3120 (s), 1735 (s), 1630 (s), 1600 (m), 1550 (s), 1350 (s), 1280 (s), 170 (s), 770 (s), 730 (s), 720 (s) cm⁻¹; 6 (400 MHz) 0.93 (1.37~1.64 (5H, m), 1.68 (1H, dd, J=3, 14 Hz), 1.70 (1H, ddd, J=5.0, 13.5, 13.5 Hz), 2.00 (1H, dddd, J=4.5, 4.5, 13.5, 13.5 Hz), 2.03 (1H, dd, J=3, 14 Hz), 2.23-2.33 (1H, m), 3.52 (1H, br.d, J=11 Hz), 3.74 (1H, dd, J=5, 11 Hz), 3.81 (1H, dd, J=11, 11 Hz), 3.86 (1H, dd, J=2.8, 11 Hz), 3.92 (1H, d, J=10 Hz, OH), 4.07~4.13 (1H, m), 4.40 (1H, dd, J=7.5, 11 Hz), 4.55 (1H, dd, J=7.0, 11 Hz), 9.14 (2H, d, J=2 Hz), 9.23 (1H, t, J=2 Hz). 25e: TLC (n-hexane-EtOAc (3:1, x3)] Rf 0.58; its physical and spectral data were described later.

Thirdly eluted fractions (10:1~5:1) gave le and 23e (730 mg, 40 %) as crystals, TLC [n-hexane-EtOAc (3:1)] Rf 0.13. This mixture was further purified by fractional recrystallization, and the mother liquor was also purified by prep TLC (nhexane-EtOAc (3:1, x 5) to afford 23e (123 mg, 17 %) as a less polar isomer and 1e (509 mg, 70 %) as a more polar isomer. 23e: TLC [n-hexane-Etchc (3:1, x4)] Rf 0.40; m.p. 171.5~173.0° (needles); [a] $^{18}_{0}$ +136° (c=0.63); vmax 3460 (s), 3120 (w), 2990 (m), 1750 (s), 1630 (m), 1610 (w), 1550 (s), 1350 (vs), 1280 (s), 1170 (s), 735 (m), 730 (m), 725 (m) cm⁻¹; 6 (400 MHz) 0.93 (3H, t, J-7.3 Hz), 1.19 (2H, dq, J-7.3, 5.3 Hz), 1.35~1.55 (3H, m), 1.54~1.64 (1H, m), 1.66 (1H, ddd, J-4.5, 13.0, 13.0, Hz), 1.74~1.87 (1H, br.s, OH), 1.94 (1H, dd, J=5.6, 13.0 Hz), 2.00 (1H, dddd, J=4.5, 4.5, 13.0, 13.0 Hz), 2.32~2.39 (1H, m), 3.44 (1H, ddd, J=11.5, 3.0, 3.0 Hz), 3.79 (1H, dd, J=3.0, 11.5 Hz), 3.84 (2H, seemingly d, J=2.0 Hz), 4.43 (1H, ddd, J=5.6, 5.6, 11.2 Hz), 4.62 (1H, dd, J=9.8, 11.0 Hz), 4.84 (1H, dd, J=4.0, 11.0 Hz), 9.16 (2H, d, J=2.0 Hz), 9.23 (1H, t, J=2.0 Hz). (Found: C, 54.16; H, 5.54; N, 6.50. Calc for C₁₉H₂₄O₉N₂: C, 53.77; H, 5.70; N, 6.60 \). 1e: TLC (n-hexane-EtOAc (3:1, x 3)] Rf 0.34; m_pp. 147-148° (fine needles); $\{\alpha\}\]^{8}$ -113° (c=2.1 (3H, t, J=7.5 Hz), 1.07-1.25 (2H, m), 1.41 (1H, dddd, J=3.5, 12.0, 12.0, 12.0 Hz), 1.42~1.61 (4H, m), 1.61~1.72 (1H, m), 1.75 (1H, ddd, J=3.5, 3.5, 13.0, Hz), 1.89 (1H, dd, J=5.6, 12.5 Hz), 2.31~2.39 (1H, m), 3.24 (1H, dd, J=11.0, 11.0 Hz), 3.56 (1H, ddd, J=2.0, 4.5, 11.0 Hz), 3.79 (1H, dd, J-2.0, 12.0 Hz), 3.83 (1H, dd, J-2.0, 12.0 Hz), 4.44 (1H, ddd, J-5.6, 5.6, 11.2 Hz), 4.62 (1H, dd, J=9.8, 11.0 Hz), 4.84 (1H, dd, J=4.0, 11.0 Hz), 9.16 (2H, d, J=2.0 Hz), 9.23 (1H, t, J=2.0 Hz). (Found: C, 54.11; H, 5.72; N, 6.50. Calc for C₁₉H₂₄O₉N₂: C, 53.77; H, 5.70; N, 6.60 0).

Conversion of (3s,4s,6R,9R) and (3R,4R,6S,9R)-9-ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro[5.5]undecane bis (3',5'dinitrolbenzoate 1d and 23d to 1e and 23e. A mixture of 1d and 23d (487 mg, 0.79 mmol) was treated with solid R₂O₃ (487) mg, 0.79 mmol) in MeOH (5 ml)-THP (2.5 ml) at room temp. The crude product was treated with DNB-Cl (194 mg, 0.84 mmol) in C5H5N (4 ml) and CH₂Cl₂ (1 ml) followed by chromatographic purification [SiO₄ (9 g), C₆H₆-EtOAc (25:1~6:1)] to give le and 23e [137 mg, 41 % from 1d and 23d]. This was further purified by fractional recrystallization and the mother liquor was also purified by prep TLC (n-hexane-EtoAc (3:1, x 3)) to afford 23e (26 mg, 19 %) and 1e (62 mg, 45 %).

Conversion of $(1c + 2c + 24c)$ to 1e and (2e8 + 24e). A mixture of 1c, 2c and 24c (1.10 g, 3.44 mmol) was treated with Na (0.3 g, 13 mmol) in THF (3 ml) and lig. NH₃ (~30 ml) as usual. The crude product (0.86 g) was treated with DNB-Cl (796 mg, 3.5 mmol) in C5H5N (14 ml) followed by chromatographic purification over SiO₂ (20 g, C₆H₆-EtOAc (40:1-5:1)) to give a mixture [550 mg, 38 % from (1c + 2c + 24c)] of 1e, 2e and 24e. This was fractionally recrystallized from n-hexane-EtOAc to give pure le (93 mg, 17 %). A portion (21 mg) of the mother liquor was purified by prep TLC [n-hexane-EtOAc (4:1, x 20)] to give further amount of pure le (3 mg) and a mixture (12 mg) of 2e and 24e; vmax 3470 (s), 3130 (s), 1730 (s), 1635 (s), 1600 (m), 1550 (s), 1345 (s), 1280 (s), 1170 (s), 775 (m), 735 (s), 725 (s) cm⁻¹; 6 (100 MHz) 0.93 (3H, t, J=6.5 Hz), 1.05~2.20 (11H, m), 3.32~3.86 [total 2H; 3.50 (dd, J=12.0, 12.0, 2e), 3.57 (d, J=10.0, 24e), 3.77 (dd, J=3.0, 10.0 Hz, 24e), 3.76~3.86 (m, 2e)), 3.85 (1H, dd, J=10.0, 10.0 Hz), 3.93 (1H, dd, J=4.7, 10.0 Hz), 4.22 (1H, ddd, J=5.1, 13.0, 13.0 Hz), 4.55, 4.62, 4.63 and 4.84 (total 2H, each dd, J=5.1, 11.0; 5.1, 10.0; 4.7, 10.0; 4.4, 11.0 Hz), 9.15 and 9.18 (total 2H, each d, J=2.0 Hz), 9.26 (1H, t, J=2.0 Hz). MS: m/z 423 (M⁺-1), 406, 341, 338, 213, 129, 126.

Conversion of (2e + 23e + 24e + 25e + 26e) to (2d + 24d), 25e and (2e + 24e). A mixture of 2e, 23e, 24e, 25e and 26e (507 mg, 1.19 mmol) was treated with K₂O₃ (0.17 g, 1.2 mmol) in MeOH (3 ml)-CHCl₃ (0.5 ml) at room temp followed by chromatographic purification over SiO_2 (4 g, n-hexane-EtOAc (3:1-2:1)) to give 237 mg (86 t) of an oil. This was treated with concernent RCl-H₂O-THF (1:5:20, 4 ml) for isomerization, cf.8 The residue (227 mg) was treated wi C5H₅N (5 ml) followed by chromatographic purification over SiO₂ (15 g). Firstly eluted fractions $(C_6H_6-\text{EtORC } (30:1\sim20:1))$
gave a mixture (99 mg, 16 t) of 2d and 24d. Analytical sample [needles from n-hexane-EtOAc (2.9° (c=0.83), vmax 3120 (w), 1735 (s), 1550 (s), 1350(s), 1280 (s), 1165 (s), 735 (s), 725 (s) cm⁻¹, 6 (100 MHz) 0.91 (~1,2H, t, J=6,1 Hz, 2d), 0,95 (-1,8H, t, J=6,1 Hz, 24d), 1,05~2,13 (8H, m), 2,31 (1H, dd, J=5,8, 10,2 Hz), 2,56 (1H, dddd, J-4.7, 5.8, 11.0, 11.0 Hz), 3.27 (~0.4H, dd, J=10.6, 10.6 Hz, 2d), 3.54 (~0.6H, br.d, J=11.5 Hz, 24d), 3.63 (~0.4H, br.d, J=10.6 Hz, 2d), 3.83 (1H, dd, J=11.0, 11.0 Hz), 3.85 (~0.6H, br.d, J=11.5 Hz, 24d), 4.03 (1H, dd, J=5.8, 11.0 Hz), 4.49 (2H, br.d, J=4.7 Hz), 5.65 (1H, ddd, J=5.1, 11.0 Hz), 9.12 and 9.14 (total 4H, each d, J=2.0 Hz), 9.25 (2H, t, J=2.0 Hz). (Found: C, 51.12; H, 4.23; N, 8.96. Calc for C₂₆H₂₆O₁₄N₄: C, 50.49; H, 4.24; N, 9.06 %).

Secondly eluted fractions (20:1-10:1) gave 25e (50 mg, 12 %). Analytical sample (plates from n-hexane-EtOAc): m.p. 128.8-129.2°; [a) $\frac{24}{3}$ -77.7° (c=0.33); vmax 1730 (s), 1630 (w), 1545 (s), 1350 (s), 1290 (s), 1165 (m), 730(m), 725 (m) cm⁻¹;

6 (400 MHz) 0.90 (3H, t, J=7.3 Hz), 1.09-1.29 (2H, m), 1.43 (1H, dddd, J=3.2, 12.5, 12 (1H, dd, J=3.0, 14.5 Hz), 2.01 (1H, dd, J=3.0, 14.5 Hz), 2.23~2.33 (1H, m), 3.33 (1H, dd, J=11.0, 11.0 Hz), 3.66 (1H, ddd, J-2.0, 4.0, 11.0 Hz), 3.73 (1H, dd, J=5.0, 11.5 Hz), 3.79 (1H, dd, J=11.5, 11.5 Hz), 3.92 (1H, d, J=10.0 Hz, OH), 4.07~4.14 (1H, m), 4.39 (1H, dd, J=7.5, 11.0 Hz), 4.56 (1H, dd, J=7.0, 11.0 Hz), 9.14 (2H, d, J=2.0 Hz), 9.23 (1H, t, J=2.0 Hz). (Found: C, 54.19; H, 5.59; N, 6.56. Calc for C₁₉H₂₄O₉N₂: C, 53.77; H, 5.70; N, 6.62 %).

Thirdly eluted fractions (10:1~5:1) gave a mixture (210 mg, 50 %) of 2e and 24e. A small amount (100 mg) was further purified by SiO₂ chromatography (3 g, C_GH_G-EtOAc (20:1~8:1)] followed by prep TLC [n-hexane-EtOAc (3:1 x 3)] to give 24e as a viscous oil (61 mg, 61 %) and 2e (21 mg, 21 %) as crystals. 24e: [a] δ^4 +21.4° (\sim 0.51); vmax 3450 (m), 3120 (m), 1735 (s), 1635 (m), 1600 (s), 1550 (s), 1350 (s), 1280 (s), 1170 (m), 1080 (m), 735 (s), 725 (s) cm⁻¹; 6 (400 MHz) 0.92 (3H, t, J=7.3 Hz), 1.36~1.44 (2H, m), 1.45 (1H, dd, J=10.5, 12.5 Hz), 1.45~1.54 (3H, m), 1.54~1.63 (1H, m), 1.69 (1H, ddd, J=4.5, 13.5, 13.5 Hz), 1.99 (1H, dddd, J=4.5, 4.5, 13.5, 13.5 Hz), 2.03~2.09 (1H, m), 2.12 (1H, dd, J=5.0, 12.5 Hz), 3.43 (1H, br.d, J=11.5 Hz), 3.62 (1H, dd, J-11.2, 11.2 Hz), 3.78 (1H, dd, J=3.0, 11.5 Hz), 3.86 (1H, dd, J=5.0, 11.2 Hz), 3.97 (1H, ddd, J-5.0, 10.5, 10.5 Hz), 4.60 (1H, dd, J-6.0, 11.5 Hz), 4.64 (1H, dd, J-4.0, 11.5 Hz), 9.14 (2H, d, J-2.0 Hz), 9.24 (1H, t, J=2.0 Hz). (Found: C, 53.76; H, 5.75; N, 6.42. Calc for C₁₉H₂₄O₉N₂: C, 53.77; H, 5.70; N, 6.60 %). 2e (fine needles from n-hexane-EtOAc): m.p. $104-104.2^{\circ}$, α 1 β ⁴ -29.8° (α e0.26), \overline{v} max 3460 (m), 3130 (w), 1730 (s), 1635 (m), 1550 (s), 1350 (s), 1280 (s), 1045 (s), 735 (m), 725 (m) cm⁻¹; 6 (400 MHz) 0.89 (3H, t, J=7.5 Hz), 1.07~1.24 (2H, m), 1.41 (1H, dddd, J=3.0, 12.0, 12.0, 12.0 Hz), 1.42~1.49 (1H, m), 1.50 (1H, dd, J=11.0, 12.2 Hz), 1.55 (1H, ddd, J=4.0, 13.0, 13.0 Hz), 1.62~1.68 (1H, m), 1.74 (1H, ddd, J=3.0, 3.0, 13.0 Hz), 1.93 (1H, br.s, OH), 2.00~2.11 (1H, m), 2.07 (1H, dd, J=5.0, 12.2 Hz), 3.23 (1H, dd, J=11.0, 11.0 Hz), 3.55 (1H, ddd, J=2.0, 4.5, 11.0 Hz), 3.58 (1H, dd, J=11.5, 11.5 Hz), 3.84 (1H, dd,
J=5.0, 11.5 Hz), 3.98 (1H, ddd, J=5.0, 11.0, 11.0 Hz), 4.58 (1H, dd, J=7.0, 11.0 Hz), 4.66 (1H, dd, J= (2H, d, J=2.0 Hz), 9.24 (1H, t, J=2.0 Hz). (Found: C, 53.82; H, 5.67; N, 6.63. Calc for C19H24O9N2: C, 53.77; H, 5.70; N, 6.60 \bullet).

(3R,4S,6R,9R)-9-Ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro[5,5]undecane [(-)-talaromycin A] la and (3S,4S,6R,9R)-9-
Ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro[5,5]undecane [(-)-talaromycin B] 2a. le (100 mg, 0.24 m with K₂CO₃ (36 mg) in MeCH (1 ml)-THF (0.5 ml) at room temp followed by chromatographic purification (SiO₂ (0.8 g), nhexane-EtOAc (4:1~2:1) followed by Et₂0] to give talaromycin A la (57 mg, quantitative), Analytical sample (hygroscopic needles from n-pentane-Et₂0!: m.p. 19-20°; [a]²⁴ -146° (c=0.46) [lit.⁸ [a]²⁰ -110.2° (c=0.83, CHCl3), 91-93 & a.e.; lit.⁹
[a]²⁶ -124.9° (c=1.11, CHCl₃)]; ORD (c=8.9 x 10⁻², MeCH; t=25°) [[a] (λ , nm)] -1 (350), -786° (300), -1382° (250); vmax 3420 (s), 2970 (s), 2950 (s), 2920 (s), 2880 (s), 1470 (m), 1460 (m), 1450 (m), 1380 (m), 1340 (w), 1290 (w), 1260 (w), 1250 (w), 1240 (m), 1215 (m), 1185 (s), 1160 (s), 1150 (m), 1125 (w), 1105 (s), 1075 (vs), 1045 (s), 1020 (s), 995 (vs), 970 (m), 940 (w), 925 (w), 890 (w), 860 (s), 805 (m), 770 (m), 760 (m), 695 (w) cm⁻¹. vmax (CHCl₃) 3680 (w), 3620 (s), 3450 (s), 3020 (s), 2980 (s), 2950 (s), 2900 (s), 1465 (s), 1445 (s), 1385 (s), 1330 (m), 1290 (m), 1265 (s), 1180 (s), 1160 (s), 1130 (m), 1100 (vs), 1070 (vs), 1040 (vs), 995 (vs), 955 (m), 945 (m), 930 (w), 910 (w), 885 (w), 865 (s), 830 (w), 670 (s), 585 (m), 570 (m) cm⁻¹. Its IR spectrum as CHCl₃ soln was identical with that provided by Prof. A. B. Smith, III. & (400 MHz) 0.88 (3H, t, J=7.5 Hz), 1.06-1.24 (2H, m), 1.38 (1H, dddd, J=4.0, 13.0, 13.0, 13.0 Hz), 1.36~1.48 (1H, m), 1.51 (1H, ddd, J=4.2, 13.0, 13.0 Hz), 1.59~1.66 (1H, m), 1.67~1.73 (1H, m), 1.72 (1H, dd, J=11.0, 13.0 Hz), 1.89 (1H, ddd, J=1.0, 5.0, 13.0 Hz), 2.11~2.18(1H, m), 2.25 (2H, br.s, OHx2), 3.19 (1H, dd, J=11.0, 11.0 Hz), 3.52 (1H, ddd, J=2.0, 4.5, 11.0 Hz), 3.58 (1H, dd, J=1.5, 12.0 Hz), 3.75 (1H, dd, J=2.8, 12.0 Hz), 3.80 (1H, dd, J-5.0, 11.0 Hz), 4.21 (1H, dd, J-9.0, 11.0 Hz), 4.41 (1H, ddd, J-5.0, 5.5, 11.0 Hz). Its ¹H NMR spectrum was identical with that reported.¹ ¹³C NNR 6 11.1, 24.9, 25.2, 35.3, 37.0, 40.4, 41.3, 60.9, 61.7, 65.4, 67.1, 97.2. The 6 value of the spiro center (97.2) was in good accord with that reported (97.2).¹ MS^{Cf.5b}: m/z 231 (M⁺+1 147, 146, 145, 144, 143, 129, 127, 126, 125. HR-MS: m/z 230.1525 (M⁺). Calc for C₁₂H₂₂O₄: 230.1518. (Found: C, 61.89; H, 9.65. Calc for C_1 2H₂O₄+(H₂O)/9: C, 62.08; H, 9.66 %).
1a (54.4 mg) was treated with Amberlyst[®]-15 (10 mg) in MeOH (1 ml) followed by chromatographic purification (SiO₂ (1

g), n-bexane-BtOAc (3:1-2:1) followed by Et₂0] to give 2a (42 mg, 78 ℓ from 1e) as a colorless viscous oil, n_0^{25} 1.4828; $\lceil \alpha \rceil$ ³ -B9.1° (c=0.48) [lit.⁹ $\lceil \alpha \rceil$ ³ -B4.1° (c=0.46, CHCl₃), 91~93 8 e.e.11 ORD (c=5.6 x 10⁻², NeOH; t=24°) [[α] (λ , nm)] -357° (600), -393° (500), -473° (400), -508° (350), -625° (300), -648° (250); vmax 3370 (s), 2980 (s), 2950 (s), 2900 (s), 1470 (E), 1450 (s), 1295 (w), 1265 (w), 1245 (w), 1215 (ml, 1185 (a), 1160 (s), 1130 (a), 1090 (61, 1070 (E), 1040 (s), 1010 (81, 990 (a), 965 (w), 955 (w), 930 (w), 890 (a), 875 (81, 825 (w), 815 (w), 795 (a), 765 (81, 700 (w), 690 (w) **cm-', vmex** Kxl3) 3670 (w), 3620 (s), 3470 (81, 3010 (81, 2970 (s), 2950 (a), 2690 (8). 1470 (8). 1450 (6). 1440 (a), 1385 (8). 1340 (w), 1330 (w), 1295 (m), 1260 (s), 1240 (s), 1225 (s), 1180 (a), 1155 (s), 1140 (s), 1125 (s), 1085 (s), 1070 (s), 1040 (s), 1010 (vs), 990 (s), 970 (m), 955 (m), 930 (m), 890 (a), 870 (s), 665 (s), 615 (m), 575 (m), 565 (m). Its IR spectrum as CHCl₃ soln was virtually identical with that provided by Prof. A. B. Smith, III. 6 (400 MHz) O.88 (3H, t, J-7.5 Hz), 1.06-1.24 (ZH, m), 1.40 UH, dddd, J-3.6, 13.0, 13.0, 13.0 Hz), 1.45 (lH, dd. J-11.0, 12.5 Hz), 1.39-1.50 (1H. m), 1.54 (1H. ddd, J-4.0, 13.0, 13.0 Hz), 1.58-1.65 UH, m), 1.71 (lH, ddd, J-2.5, 3.6, 13.0 Hz), 1.78-1.91 (1H. m). 1.99 (iii, dd, J-5.0, 12.0 Hz), 2.80 (ZH, br.s, C4tx2), 3.20 (lH, dd, J-11.0, 11.0 Hz), 3.32 UH, dd, J-11.0, 11.0 Hz), 3.51 (1H. ddd, J-2.0, 4.5, 11.0 Hz), 3.59 (lH, dd, J-5.0, 11.0 He), 3.70 (ZH, d, J-6.0 Hz), 4.05 (lH, ddd, J-5.0, 10.5, 11.0 HZ). rta 1~ **NPIR** spectrum wa8 identical with that reported.1*2*5b l3 C NWR 6 11.1, 24.8, 25.2, 35.2, 36.7, 44.1, 45.4, 60.4, 62.9, 65.3, 67.2, 97.2. Its ¹³C NMR spectrum was identical with that of (±)-2a reported previously.³⁰ MS³⁰: <u>m</u>/<u>z</u> 231 (M⁺+1), 230 (M⁺), 212, 200, 157, 153, 147, 146, 145, 144, 143, 129, 127, 126, 125. HR-MS: m/z 230.1525 (M⁺). Calc for C12H220q: 230.1518.

CD spectra of talaromycin A dibenzoate 1g, talaromycin B dibenzoate 2g and (3R,4S,6R,9R)-9-ethyl-4-hydroxy-3-hydroxymethyl- $1/7$ -dioxaspiro[5.5]undecane dibenzoate 259. Ig: (-6.7×10^{-2} , EXOH, t=25°) [ΔE (λ , nm)] +9.5 (238), 0 (227.5), -6.3 (221); (σ 5.7 x 10⁻², n-hexane, t-25^o) [$\Delta \epsilon$ (λ , nm)] +11.5 (233), 0 (224), -4.3 (219). 2g: (σ ⁹A x 10⁻², EtcH, t-25^o) [$\Delta \epsilon$ (λ , nm)] +3.3 (232), +0.57 (223); (c=3.0 x 10⁻², n-hexane, t-25^o) [Δε (λ, nm)] -0.4 (248), 0 (240), +1.0 (230). 25g: (c=9.3 x 10⁻², B tOH, $t=25^{\circ}$) [$\Delta\varepsilon$ (λ , nm) -7.3 (238), 0 (228), +3.1 (221). Its CD spectrum was antipodal to that of (25,4S,5S)-13c.

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