SYNTHESIS OF (-)-TALAROMYCINS A AND B⁺

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Abstract -- Highly enantiomerically pure (-)-talaromycins A and B [(3R,4S,6R,9R)- and (3S,4S,6R,9R)-9-ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro[5.5]undecane] were synthesized starting from chiral building blocks of microbial origin.

(-)-Talaromycins A 1a and B 2a were fungal toxins isolated by Lynn and co-workers in 1982 from <u>Talaromyces stipitatus</u>.^{1,2} The toxicity of these spiroacetals may be due to their ability to block outward potassium fluxes, thus leading to muscle dysfunction.¹ Their unique spiroacetal structures 1a and 2a attracted much attention of synthetic chemists. Thus thermodynamically more stable (±)-talaromycin B 2a with an eq CH₂OH group was synthesized by Schreiber <u>et al</u>.,³ Kozikowski <u>et al</u>.,⁴ Kocienski <u>et al</u>.⁵ and Kay <u>et al</u>.⁶ The less stable (±)-talaromycin A 1a with an ax CH₂OH group was later synthesized by Schreiber <u>et al</u>.⁷ As to the synthesis of the naturally occurring enantiomers of talaromycins, there exist only two reports.^{8,9} Smith and Thompson was the first to synthesize (-)-talaromycins A and B in 1984.⁸ The second synthesis of (-)-talaromycin A was reported by Midland and Gabriel in 1985.⁹ As an extension of our continuing efforts to synthesize enantiomerically pure spiroacetals of insect origin,^{10,11} we initiated a project to synthesize both (-)-talaromycins A and B in highly enantiomerically pure state.

Our synthetic plan is shown in Fig. 1. Chiral building blocks of microbial origin are employed as our starting materials. For the construction of the spiroacetal system, a Wittig reaction^{12~14} between A and B is to be employed. The phosphorane A can be prepared from C, which in turn is the product of yeast reduction^{15~17} of D. Dimethyl 3-oxopentanedioate E is the starting material for D. The aldehyde B is to be synthesized <u>via</u> F from ethyl (<u>S</u>)-3-hydroxybutanoate G, the product of yeast reduction of ethyl acetoacetate.¹⁸ Execution of the above plan, which culminated to the synthesis of crystalline (-)-talaromycin A 1a, will be detailed below.

The first phase of our work as shown in Fig. 2 was the synthesis of a phosphonium salt 14, the precursor to the Wittig reagent A. Reduction of 3 (=E) with $NaBH_4$ gave 4a. After protecting the OH group of 4a as a THP ether, the resulting 4b was reduced with LAH

[†]Synthetic Microbial Chemistry — XIII. Part XII, T. Kitahara, H. Kurata, T. Matsuoka and K. Mori, <u>Tetrahedron</u> 41, 5475 (1985). The experimental part of this work was taken from the forthcoming doctoral dissertation of M. I. (March, 1987).



Fig. 1. Synthetic plan.

to give 5a. This was treated with p-TsOH in MeOH to give a triol 5b. Acetonide 6 was prepared from 5b in the usual manner. Oxidation of 6 with pyridinium chlorochromate $(PCC)^{19}$ in the presence of MS 3\AA^{20} furnished aldehyde 7. When a soln of 7 and p-TsOH in CH_2Cl_2 -EtOH was heated, intramolecular acetalization took place to give a stereoisomeric mixture of 8 in 64 % yield from 7 or in 33 % overall yield from 3 in 7 steps. Although a synthesis of 4-hydroxy-2-methoxytetrahydropyran was known,²¹ the present multi-step route furnished 8 in better overall yield. The alcohol 8 was oxidized with pyridinium dichromate $(PDC)^{22}$ in the presence of MS 3\AA^{20} to give 9 in 72 % yield. Ethoxycarbonylation of 9 could not be achieved under the conventional condition employing NaH and $CO(OEt)_2$. By the method of Mander and Sethi,²³ however, a CO_2Et group was successfuly introduced at C-5 of 9. Thus 9 was treated with LDA and $NCCO_2Et^{24}$ to give a mixture of two regioisomeric β -keto esters 10 and 11 in a ratio of 10:1. This was purified by SiO₂ chromatography to give pure 10 in 55 % yield.

The next step was the crucial microbial reduction of 10. Reduction of the β -keto ester 10 was tried with <u>Saccharomyces bailii</u> KI 0116, <u>Pichia terricola</u> KI 0117 and <u>Saccharomyces cerevisiae</u> (baker's yeast).¹⁸ <u>S. bailii</u> and <u>P. terricola</u> were found to be unsuitable for this reduction giving almost no β -hydroxy ester, although <u>S. bailii</u> was an excellent organism to reduce a β -keto ester related to PGI₂.²⁵ Reduction of 10 was therefore carried out with baker's yeast. An emulsion of 10 in dilute Triton X-100 was added to a suspension of briskly fermenting baker's yeast at pH 8 (phosphate buffer) in the presence of sucrose. The product obtained in 81 % yield was a diastereomeric mixture at C-2 of hydroxy ester 12a. The diastereomeric ratio was later determined as 63:37 by analyzing 13c (vide infra). The diastereomers of 12a were partially separated by SiO₂ chromatography to give the less polar isomer (major product), the more polar isomer (minor product) and their mixture. In the NMR spectrum of the less polar isomer of 12a, a signal



Fig. 2. Synthesis of the phosphonium salt 14.

due to an eq H at C-2 (δ 4.78, 1H, dd, <u>J</u>=3 and 3 Hz) and that due to an ax H at C-5 (δ 2.53, 1H, ddd, \underline{J} =10, 4 and 2 Hz) were observed. In all of the published examples of the yeast reduction of cyclic β -keto esters, formation of <u>cis</u>- β -hydroxy esters were observed with (S)-configuration of the OH group.15 - 17, 25, 26 The less polar isomer of 12a was therefore assumed to be (2<u>S</u>,4<u>S</u>,5<u>R</u>)-12a, whose enantiomeric purity was estimated to be 43 % e.e. by the HPLC analysis of the corresponding (\underline{R}) - and (\underline{S}) - α -methoxy- α -trifluoromethylphenylacetates (MTPA esters²⁷), $(2\underline{s},4\underline{s},5\underline{R})$ -12b and 12c. The more polar and minor isomer of **12a** exhibited in its NMR spectrum a signal due to an eq H at C-2 (δ 4.71, 1H, dd, <u>J</u>=4.4 and 3.6 Hz) and that due to an eq H at C-5 (δ 2.62, 1H, ddd, J=4.4, 4 and 3.2 Hz). Assuming the (S)-configuration for 4-OH, this isomer was thought to be (2R, 4S, 5R)-12a. Determination of the enantiomeric purity of $(2\underline{R},4\underline{S},5\underline{R})-12a$ was rather troublesome, because its MTPA ester (2R,4S,5R)-12b showed no good separation of the diastereomers when analyzed by HPLC. The minor isomer $(2\underline{R},4\underline{S},5\underline{R})$ -12a was therefore treated with <u>p</u>-TsOH in EtOH to effect equilibration at C-2 to generate $(2\underline{S},4\underline{S},5\underline{R})-12a$, whose MTPA ester $(2\underline{S},4\underline{S},5\underline{R})-12b$ was known to exhibit good diastereomer separation by HPLC. The HPLC analysis of the MTPA esters derived from the above described equilibrated mixture showed the generated $(2\underline{S}, 4\underline{S}, 5\underline{R})$ -12a to be of 100 % e.e. Accordingly the diastereomeric mixture of 12a was of 64 % e.e. [(43 x 0.63) + (100 x 0.37) = 64] with regard to the chiral centers at C-4 and C-5.

To secure a supporting evidence for the assumed absolute configuration of the two isomers of 12a, they were converted to the corresponding dibenzoates 13c and their CD spectra were measured. To prepare 13c, 12a (diastereomeric mixture) was first acetylated with Ac_2O to 12d. LAH reduction of 12d gave 13a in 97 % yield from 12a. Direct reduction of 12a with LAH afforded an inferior result. Benzoylation of 13a yielded dibenzoates 13c as a diastereomeric mixture at C-2 in a ratio of 63:37 as analyzed by HPLC. The diastereomers of 13c were separated by prep TLC to give the less polar isomer (21 % recovery) and the more polar isomer (52 % recovery). ¹H NMR spectra of these two isomers revealed that the former possessed an ax H at C-4 (δ 5.70, 1H, ddd, <u>J</u>=7.3, 4.4 and 4.1 Hz), while the latter had an eq H at C-4 (δ 5.47, 1H, ddd, <u>J</u>=5.8, 5.8 and 5.8 Hz). In both of them the C-2 H was in eq orientation (<u>J</u>=3.3~4.9 Hz) like in the starting β -hydroxy esters 12a. Application of the exciton chirality method^{28,29} to these dibenzoate isomers provided a supporting evidence concerning their absolute configuration. The exciton-split CD spectrum of the less polar isomer of 13c in EtOH showed a positive first Cotton effect at 237 nm ($\Delta\epsilon$ +11) and a negative second Cotton effect at 220 nm ($\Delta\epsilon$ -2.8). The more polar isomer of 13c also exhibited a positive first Cotton effect at 238 nm ($\Delta\epsilon$ +5.8) and a negative second Coton effect at 220 nm ($\Delta\epsilon$ -1.0). Comparison of these data with the CD spectra of some triterpene dibenzoates of similar structural feature²⁸ supported the assumed (4<u>S</u>)-configuration of the dibenzoates 13c as depicted in Fig. 3. The less polar isomer was therefore (2<u>R</u>,4<u>S</u>,5<u>S</u>)-13c and the more polar one was (2<u>S</u>,4<u>S</u>,5<u>S</u>)-13c.



Fig. 3. CD spectra of (2R,4S,5S)-13c and (2S,4S,5S)-13c.

Having clarified the absolute configuration and enantiomeric purity of 13a, the remaining task was its conversion to 14. Treatment of a diastereomeric mixture of 13a with PhCH₂Br and NaH in the presence of $(\underline{n}-Bu)_4NI$ in THF^{cf.30} gave the corresponding dibenzyl ether 13b in 80 % yield. The desired phosphonium salt 14 was prepared in quantitative yield from 13b by heating it with Ph₃PH⁺BF₄⁻ in MeCN.^{31,32} The overall yield of 14 from 3 was 8 % in 13 steps.

The second phase of our project as shown in Fig. 4 was the synthesis of the aldehyde **20** (=**B**), the other partner of the Wittig reaction. Reduction of ethyl acetoacetate **15** with a thermophilic yeast <u>Saccharomyces bailii</u> KI 0116 gave (<u>S</u>)-**16** (96~98 % e.e.) in 84 % yield.¹⁸ The diamion derived from **16** was alkylated with allyl bromide according to Fráter to give **17a** contaminated with its <u>syn</u>-isomer (<u>anti:syn</u>=96:4) in 85 % yield.³³ The diastereomeric ratio was determined by capillary GLC using a diastereomeric mixture of ethyl



Fig. 4. Synthesis of the aldehyde 20.

(±)-2-(1'-hydroxyethyl)pent-4-enoate (anti:syn=51:49) as a reference sample. Conversion of 17a to 20 was lengthy but straightforward. The OH group of 17a was protected as THP ether. Reduction of the resulting 17b with LAH gave an alcohol 18a. This was benzylated to 18b, and the THP protective group was removed from 18b to give 18c. The corresponding 3,5-dinitrobenzoate (DNB) 18d was crystalline, and could be purified by recrystallization to give diastereomerically and enantiomerically pure 18d. Treatment of 18d with KOH gave pure 18c, whose diastereomeric purity as 100 % was checked by capillary GLC. The corresponding (R)-MTPA ester 18e exhibited a single peak upon HPLC analysis proving the high enantiomeric purity (100 % e.e.) of 18c. As the task of the OH group to control the absolute configuration of the new chiral center was over, it was removed by tosylation to 18f followed by its reduction with LiBEt₃H. 34 The benzyl (Bn) protective group of 19a was then replaced by the THP group to give 19c by reducing 19a with Na/liq NH3 and protecting the OH group of 19b. Finally, the Lemieux-Johnson oxidation of 19c with OsO_4 -NaIO₄ gave (R)-20 in 34 % overall yield from 15 in 13 steps. By switching the protective group from Bn to THP, the construction of the spiroacetal system became less complicated due to the more facile deprotection of the THP group compared with that of the Bn group.

The final stage of the synthesis as shown in Fig. 5 was the coupling of the two building blocks 14 and 20 by the Wittig reaction to give the desired 1,7-dioxaspiro[5.5] undecane system. Treatment of the phosphonium salt 14 with n-BuLi in THF-HMPA generated a deep red phosphorane 21. This was condensed with the aldehyde 20 to give a cyclic enol ether 22. Acid treatment (conc HCl:H₂O:THF=1:5:20) of crude 22 gave a mixture of spiroacetals. As one of the building blocks was optically impure (64 % e.e.), formation of four spiroacetal dibenzyl ethers (1b, 23b, 25b and 26b) were expected. However, the products obtained after SiO₂ chromatography were a mixture of dibenzyl ethers 1b and 23b (46 % from 14) and also a mixture of 12-monobenzyl ethers 1c, 2c and 24c (35 % from 14). The unexpected formation of 12-monobenzyl ethers could be rationalized as shown in Fig. 5 by the sequence $22 \rightarrow \alpha \rightarrow \beta \rightarrow \gamma \rightarrow 1c + 2c + 24c$ involving (i) olefin isomerization (ii) retro-Michael-like elimination of PhCH₂OH (iii) addition of H₂O and (iv) spiroacetal formation. Under the acid condition employed, the above process competed to some extent with the desired hydrolysis of the THP group followed by spiroacetalization. The elimination-addition mechanism as above allowed the formation of 2c and 24c, where the OH group and the CH_OBn group were in trans-relationship in contrast to the original cis-relation-



Fig. 5. Synthesis of Talaromycins A and B.

ship in 21. The structures 1b, 23b, 1c, 2c and 24c were deduced on the basis of the NMR analysis of thir 4-benzoates (1f, 2f and 24f) or that of their 12-mono DNB derivatives 1e, 2e and 23e (see Experimental). Other two possible isomers could not be isolated from the reaction mixture.

Crystalline 12-mono DNB derivative 1e of (3R,4S,6R,9R)-talaromycin A 1a was derived from the mixture of the dibenzyl ethers 1b and 23b in the following manner. The mixture (2.35 g) of 1b and 23b was reduced with Na/liq NH₃ to give a mixture of 1a and 23a in 75 % yield. Treatment of the mixture of 1a and 23a with 3,5-dinitrobenzoyl chloride (DNBCl) in pyridine in the presence of $4-(\underline{N},\underline{N}-dimethylamino)$ pyridine (DMAP) gave a complex mixture of products. This was chromatographed over SiO, to give, in the order of elution, a mixture of bis DNB esters (1d and 23d; 18 % yield), a mixture of new mono DNB esters (25e and 26e; 14 % yield), and a mixture of talaromycin A 12-mono DNB ester 1e and its isomer 23e (40 % yield). The new mono DNB esters 25e and 26e must have been generated by epimerization at the spiro-center in the course of the acylation reaction, because they could be detected by TLC monitoring of the acylation reaction. The epimerization was due to the preference of the CH₂ODNB group to adopt eq orientation. Pure talaromycin A 12-mono DNB ester 1e (509 mg), m.p. 147~148°, $[\alpha]_D^{18}$ -113° (CHCl₃), was obtained by recrystallization of the mixture (730 mg) of 1e and 23e. Pure 23e (123 mg), m.p. 171.5~173°, $[\alpha]_D^{18}$ +136° (CHCl₃), was also secured by purification with prep TLC and recrystallization. The mixture of bis DNB esters (1d and 23d) was hydrolyzed with K_2CO_3 to a mixture of 1a and 23a, which was acylated again and recrystallized to give additional amounts of 1e (62 mg) and 23e (26 mg). A further amount of 1e (93 mg) was obtained from a mixture (1.10 g) of monobenzyl ethers (1c, 2c and 24c) by reduction with Na/liq NH3, acylation with DNBCl and recrystallization of the resulting mixture of mono DNB esters 1e, 2e and 24e. In total, talaromycin A 12-mono DNB ester **1e** was obtained in 13 % overall yield from **13b** in 5 steps. From the combined mother liquor of the above-mentioned recrystallization experiments, small amounts of pure talaromycin B 12-mono DNB ester 2e, m.p. 104~104.2°, $[\alpha]_D^{24}$ ~29.8° (CHCl₃), and another mono DNB ester 25e, m.p. 128.8~129.2°, $[\alpha]_D^{24}$ -77.7° (CHCl₃), were isolated. The amount of pure **2e** was so small that its hydrolysis to talaromycin B **2a** was not attempted. In Fig. 6 is shown the ratio of four 12-mono DNB esters (1e, 23e, 25e and 26e) generated from the mixture of dibenzyl ethers (1b and 23b) as expressed in percentages. As can be seen from Fig. 6, the ratio of the isomers reflected the enantiomeric purity of the phosphorane 21.



Fig. 6. Derivation of the 12- mono DNB esters of talaromycin stereoisomers.

To complete the synthesis, talaromycin A mono DNB ester 1e was hydrolyzed with K_2CO_3 in MeOH-THF to give, in 89 % yield, the natural enantiomer of talaromycin A 1a for the first time as crystals, m.p. 19~20°, $[\alpha]_D^{24}$ -146° (CHCl₃) [lit.⁸ $[\alpha]_D^{20}$ -110.2° (CHCl₃);

lit.⁹ $[\alpha]_D^{26}$ -124.9° (CHCl₃)]. Our success in synthesizing highly pure and crystalline (-)-talaromycin A 1a may be due to the nicely crystalline nature of its 12-mono DNB ester 1e. Because of that property we were able to purify it completely. (-)-Talaromycin B 2a, $[\alpha]_D^{24}$ -89.1° (CHCl₃) [lit.⁸ $[\alpha]_D^{20}$ -84.1° (CHCl₃)], was also prepared from 1e by hydrolysis with K₂ ∞_3 followed by acid-catalyzed isomerization with Amberlyst^e-15 (H⁺-form) in MeOH in 78 % yield. The 400 MHz ¹H NMR spectra of our synthetic (-)-talaromycins A and B were in good accord with the authentic spectra of the natural products kindly provided by Prof. Lynn (360 MHz)¹ and Prof. Smith (250 and 500 MHz).⁸ The ¹³C NMR spectrum of (-)-talaromycin B dibenzoates 1g and 2g of (-)-talaromycins A and B are shown in Fig. 6. They were also



Fig. 7. CD spectra of the dibenzoates of talaromycins A and B.

identical to Prof. Lynn's authentic spectra of the dibenzoates derived from the natural products. The overall yield of 1a was 4 % from ethyl acetoacetate 15 in 18 steps or 0.9 % from dimethyl 3-oxopentanedioate 3 in 19 steps. (-)-Talaromycin B 2a was obtained in 3 % overall yield from 15 in 19 steps or in 0.8 % overall yield from 3 in 20 steps.

In summary, highly pure natural enantiomers of talaromycins A and B were synthesized in a convergent manner employing the chiral building blocks of microbial origin. The yeast reduction was again shown to be useful in organic synthesis. An enantiomerically impure intermediate such as 21 can satisfactorily be employed to afford enantiomerically pure target molecules after removing the unwanted diastereomers. Experimental importance of a crystalline intermediate in a chiral synthesis was also exemplified in the present case. The principle of 'optical enrichment'^{9,11} in the case of a chiral synthesis was applicable to the present synthesis, too.

EXPERIMENTAL

All bus and muss were uncorrected. IR spectra were measured as films for oils or as mujol mulls or KBr discs for

eolids on a Jasco IRA-102 or IRA-202 spectrometer unless otherwise stated. ¹H NMR spectra were recorded with TMS as an internal standard on a Hitachi R-24A (60 MHz) or on a JEOL JNM FX-100 (100 MHz, CDCl₃) or on a JEOL JNM GX-400 (400 MHz, CDCl₃) spectrometer. ¹³C NMR spectra were measured with TMS as an internal standard as CDCl₃ soln at 25 MHz on a Jeol JNM FX-100 spectrometer. Optical rotations were measured on a Jasco DIP 140 polarimeter as CHCl₃ soln. GRD and CD spectra were measured on a Jasco J-20C spectropolarimeter. Mass spectra were recorded on a JEOL DK-303 or on a Hitachi RMU-6M spectrometer at 70 eV. Fuji-Davison BW-820 MH was used for SiO₂ column chromatography. Merck Kieselgel 60 Art 5717 or 5744 were used for prep TLC separation. HPLC analyses were performed on Nucleosil[®] 50-5 (25 cm x 4.6 mm) by the detection at 254 nm unless otherwise stated.

<u>Dimethyl 3-tetrahydropyranyloxypentanedioste</u> **4b.** Dimethyl 3-oxopentanedioste 3 (200 g, 1.15 mol) was reduced with NaEH₄ (16.7 g, 0.44 mol) in MeOH (700 ml) in a usual manner to give crude dimethyl 3-hydroxypentanedioste **4a** (213 g, quantitative), wmax 3550 (s), 1740 (s), 1200 (s) cm⁻¹. Crude **4a** (210,3 g, ~1.14 mol) was treated with dihydropyran (138 g, 1.64 mol) and p-TsOH-H₂O (0.4 g, 2.1 mmol) in Et₂O (1.2 l) in a usual manner to give crude **4b** (296.9 g, quantitative). Analytical sample: b.p. $105 \sim 121^{\circ}/1.0$ Torr, n_{2}^{16} 1.4521; vmax 1740 (s), 1200 (s) cm⁻¹, δ (CCl₄) 1.10~1.95 (6H, m), 2.47, 2.54, 2.57 and 2.62 (total 4H, each d, J=6 Hz), 3.10~3.90 (2H, m), 3.64 (6H, s), 4.36 (1H, t, J=6 Hz), 4.72 (1H, br.s). (Found: C, 55.47; H, 7.68. Calc for Cl₂H₂O₆: C, 55.37; H, 7.75 %).

Pentane-1,3,5-triol 3-THP ether 5a. Crude 4b (129 g, 0,50 mol) was reduced with LiAlH₄ (LAH, 30 g, 0,69 mol) in Et₂O (720 ml) in a usual manner to give crude 5a (100 g, 98 % from 3); ymax 3420 (s), 1140 (s), 1080 (s), 1030 (s) cm⁻¹; & (CDCl₃) 1.00~2.40 (10H, m),3.32 (2H, s, OHx2), 3.10~4.38 (7H, m), 4.4~4.8 (1H, br.s).

 $\frac{\text{Pentane-1,3,5-triol}}{\text{manner to give crude 5a (235,7 g, ~1.15 mol) was treated with p-TsOH·H₂O (2.6 g, 14 mmol) in MeOH (1.2 1) in a usual manner to give crude 5b (146,9 g, quantitative). Analytical sample: b.p. 154~156°/1.3 Torr; <math>n_1^{18}$ 1.4655; vmax 3400 (s), 1050 (s) cm⁻¹; & (C₅D₅N-CDCl₃) 2.00 (4H, dt, J=6, 6 Hz), 4.09 (4H, t, J=6 Hz), 4.43 (1H, t, J=6 Hz), 5,86 (3H, s, OHx3). (Pound: C, 50.35; H,10.15. Calc for C₅H₁₂O₃: C, 49.98; H, 10.07 %).

<u>Pentane-1,3,5-triol</u> <u>1,3-acetonide</u> 6. Crude 5b (144 g, 1,13 mol) was treated with 2,2-dimethoxypropane (254 g, 2,4 mol) and p-TaOH+H₂O (6.5 g, 34 mmol) in acetone (1.2 l) in a usual manner to give 6 (98,3 g, b,p. $60 \times 107^{\circ}/15 \times 16$ Torr, 53 % from 3). A portion was chromatographed over SiO₂ [<u>n</u>-hexane-Et₂O (15:1-2:1)] followed by distillation in the presence of K₂OO₃ to give an analytical sample: b,p. 113-115°/17 Torr; n₁^B 1.4495, vmax 3450 (s), 1200 (s), 1100 (s), 1060 (s) cm⁻¹; s (CCl₄) 0.95×1.45 (2H, m), 1.32 (3H, s), 1.43 (3H, s), 1.63 (2H, dt, J=5, 5 Hz), 3.06 (1H, t, J=5 Hz, OH), 3.25~4.31 (4H, m). (Found: C, 60.40; H, 10.05. Calc for C_BH₁6O₃: C, 59.98; H, 10.07 %).

35-Dihydroxypentanal acetonide 7. According to the reported procedure, 19,20 6 (20 g, 125 mmol) was oxidized with pyridinium chlorochromate (PCC, 45,4 g, 211 mmol) in CH₂Cl₂ (310 ml) in the presence of MS 3Å (40 g) to give crude 7 (19,3 g, 97 %). A portion of it was chromatographed over SiO₂ [n-pentane-Et₂O (20:1~3:1)] followed by distillation under Ar to give an analytical sample; b_D. 83~87°/45 Torr; n₀¹⁸ 1.4419; vmax 2750 (m), 1730 (s), 1200 (s), 1165 (s), 1100 (s) cm⁻¹; δ (CCl₄) 1.26 (3H, s), 1.41 (3H, s), 0.80~1.90 (2H, m), 2.45 (2H, ddd, J=2, 2, 6 Hz), 2.46 (1H, dd, J=2, 6 Hz), 3.40~4.00 (2H, m), 4.00~4.63 (1H, m), 9.86 (1H, dd, J=2, 2 Hz). (Found: C, 60.31; H, 8.87. Calc for C₈H₁₄O₃: C, 60.74; H, 8.92 %).

<u>2-Ethoxytetrahydropyran-4-ol</u> 8 A soln of crude 7 (18.2 g, ~115 mmol) and p-TsOH+H₂O (0.4 g, 2.1 mmol) in 99 % EtOH (80 ml) and CH₂Cl₂ (240 ml) was stirred and heated under reflux overnight with azeotropic removal of water by use of MS 4Å. After cooling, the mixture was neutralized by the addition of solid Na₂OO₃ (4.6 g, 43.4 mmol) and stirred further for 4 h at room temp. The solid was filtered off through a pad of Celite and washed with Et₂O. The combined filtrate and washings were concentrated <u>in vacuo</u>. The residue was diluted with Et₂O, filtered through a pad of Plorisil, and the Florisil layer was washed with Et₂O. The combined filtrate and washings were concentrated <u>in vacuo</u>. The residue (15.8 g) was distilled in the presence of K₂OO₃ to give 8 (10.8 g, 64 %), bp. 92-94°/9 Torr; n_0^{A4} 1.4438; vmax 3430 (s), 1130, (s), 1060 (s) cm⁻¹; 6 (CCl₄) 1.18 (-2.4H, t, J=7 Hz), 1.25 (-0.6H, t, J=7 Hz), 1.35-2.40 (4H, m), 2.87-4.30 (6H, m), 4.52 (-0.2H, dd, J=3, 14 Hz), 4.78 (-0.8H, dd, J=3, 3 Hz). (Found: C, 57.41; H, 9.55. Calc for C₇H₁₄O₃: C, 57.51; H, 9.65 %). <u>2-Ethoxytetrahydropyran-4-one</u> 9. According to the reported procedure,^{20,22} 8 (47 g, 32 mmol) was oxidized with pyridinium

2-Ethoxytetrahydropyran-4-one 9. According to the reported procedure, 20,22 8 (4.7 g, 32 mmol) was oxidized with pyridinium dichromate (PDC, 24.2 g, 64.3 mmol) in CH₂Cl₂ (90 ml) in the presence of MS 3Å (34 g) followed by chromatographic purification [SiO₂ (80 g), <u>n</u>-hexane-Et₂O (10:1)] then distillation to give 9 (3.3 g, 72 %), bp. 104~107°/23 Torr; ng⁵ 1.4395; vmax 1730 (s), 1120 (s), 1060 (s) cm⁻¹; 6 (CCl₄) 1.21 (3H, t, J=7 Hz), 1.90~2.83 (4H, m), 3.10~4.35 (4H, m), 5.05 (1H, dd, J=3.6, 3.6 Hz). (Found: C, 58.11; H, 8.38. Calc for C7H₁₂O₃: C, 58.31; H, 8.39 %).

Ethyl 2-Ethoxy-4-oxo-5-tetrahydropyrancarboxylate 10 and ethyl 2-ethoxy-4-oxo-3-tetrahydropyrancarboxylate 11. A soln of LDA was prepared from n-BuLi (1.65 M in n-hexane, 12.4 ml, 20.5 mmol) and \underline{i} -Pr₂NH (2.9 g, 20.5 mmol) in THF (45 ml) under Ar in a usual manner. To this soln was added dropwise a soln of 9 (2.5 g, 17.3 mmol) in THF (15 ml) over 13 mn at -76° —72° and then added HMPA (12 ml) over 2 min at -76° —66°. The reaction temp was allowed to rise to -35° to give a homogeneous soln. To this was added a soln of ethyl cyanoformate²⁴ (2.2 g, 21.8 mmol) in THF (2 ml) all at once at -76° —56°. After stirring for 15 min at -76° — free extract was poured into ice-water, neutralized by the addition of dil HCl and extracted with Et₂Q. The extract was washed with brine, dried (MgSQ₄) and concentrated in varue. The residue (9.1 g) was chromatographed over SiO₂ (110 g, n-hexane-EtOAc (17:1)) followed by the purification of fractions contaminated with 11 by prep TLC (CgH₆-EtOAc (5:1)) to give 10 (total 2.04 g, 55 %). Analytical sample: b.p. 90-99°/0.45 Torr; n_{0}^{20} 1.4656; vmax 1750 (s), 1670 (s), 1635 (s), 1240 (s), 1060 (s) cm⁻¹; δ (CCl₄) 1.20 (3H, t, J=7 Hz), 1.29 and 1.36 (total 3H, each t, J=7 Hz), 1.90~2.80 (2H, m), 3.10~4.10 (2H, m), 4.17 (4H, q, J=7 Hz), 4.84 (1H, dd, J=2, 3 Hz), 5.12 (~0.27H, br.s), 11.77 (~0.53H, s), 12.12 (~0.2H, s). (Found: C, 55.18; H, 7.02. Calc for Cl₀H₁₆O₅: C, 55.54; H, 7.46 %). Further elution with the same solvent gave 11 (total 0.20 g, 5.4 %) vmax 1745 (s), 1680 (s), 1640 (w), 1565 (s), 1265 (s), 1150 (s) cm⁻¹; δ (CCl₄) 1.25 (3H, t, J=7 Hz), 1.35 (3H, t, J=7 Hz), 1.90~2.70 (2H, m), 3.20~4.10 (3H, m), 4.02 (2H, q, J=7 Hz), 4.28 (2H, q, J=7 Hz), 5.07 (1H, br.d, J=5 Hz); HS m/z 216 (H⁺), 215, 201, 171, 143, 73, 45, 43 (base peak).

Ethyl (25,45,5R)- and (2R,45,5R)-2-ethoxy-4-hydroxy-5-tetrahydropyrancarboxylate 12a. To a soln of sucrose (250 g, 730 mmol) in phosphate buffer (0.1 M, pH 8, 1.3 1) was added dry baker's yeast (110 g, Oriental Yeast Co., Ltd.) at 30°. After vigorous stirring for 10 min with aeration, to this fermenting mixture was added an emulsion of 10 (5,2 g, 24 mmol) in 0.2 Triton X-100 ag soln (50 ml), and the stirring was continued for 1 h at 30°. After the addition of Et₂O, Oelite and a small amount of Norit, the mixture was filtered. The filter-cake was washed thoroughly with EtOAc. The organic layer of the combined filtrate and washings was separated. The aq layer was saturated with NaCl and extracted with EtOAc. The combined EtOAc soln was dried (MgSO₄) and concentrated in vacuo. The residue (7.8 g) was chromatographed over SiO₂ (80 g). Firstly eluted fractions (n-hexane-EtOAc (15:1~12:1)) gave (25,45,5R)-12a (1.95 g, 37 %), ng^O 1.4486; (α) $\frac{10}{2}$ * 57.0° (c=0.51); vmax 3550 (s), 1730 (s), 1120 (s), 1100 (s), 1030 (s) cm⁻¹, 6 (100 MHz +D₂O) 1.26 (3H, t, J=7.2 Hz), 1.29 (3H, t, J=7.2 Hz), 1.87 (1H, ddd, J=3.4, 3.5, 14.2 Hz), 2.04 (1H, ddd, J=2.0, 3.7, 14.2 Hz), 2.72 (1H, ddd, J=2.9, 5.0, 12.2 Hz), 3.46 (2H, dq, J=7.2, 9.6 Hz), 3.76 (1H, ddd, J=5.0, 11.7 Hz), 3.67~3.98 (1H, m), 4.18 (1H, ddd, J=1.7, 12.2 Hz), 4.20 (2H, g, J=7.2 Hz), 4.89 (1H, bradd, J=2.0, 3.4 Hz); MR=MS m/z 217.1116 (M^{*}-H). Calc for C10H1705: 217.1076. Determination of the optical purity of (25,45,5R)-12a was as follows. (a) According to the reported procedure²⁷, (25,45,5R)-12a was converted to its $(\underline{R})^-$ and $(\underline{S})^-$ MTFA estar, $(2\underline{S},4\underline{S},5\underline{R})^-$ 12b and 12c, respectively. (b) HFLC analysis of $(2\underline{S},4\underline{S},5\underline{R})^-$ 12b: [Column, Silica-1251N, 25 cm x 4.6 mm, <u>n</u>-hexane-THF (20:1), 1 ml/min] Rt 56.6 min (28.5 %), 57.9 min (71.5 %). HFLC analysis of the ~1:1 mixture of $(2\underline{S},4\underline{S},5\underline{R})^-$ 12b and 12c under the same condition as above showed Rt 58.7 min (48.4 %) and 60.0 min (51.6 %). Therefore the optical purity of $(2\underline{S},4\underline{S},5\underline{R})^-$ 12a was determined to be 43 % e.e.

Secondly eluted fractions [n-hexane-BtOAc (10:1-6:1)] gave a mixture of (25,45,5R) and (2R,45,5R)-12a (1,60 g, 31 %). Thirdly eluted fractions [n-hexane-BtOAc (8:1)] gave (2R,45,5R)-12a (0,60 g, 11 %), nb⁹ 1,4549; [a]b⁶ -76.9° (c=0,54); vmax 3500 (s), 1735 (s), 1140 (s), 1065 (s) cm⁻¹; & (CCl4) 1,16 (3H, t, J=7 Hz), 1,28 (3H, t, J=7 Hz), 1.55-2,00 (2H, m), 2.62 (1H, ddd, J=3.2, 4.0, 4.4 Hz), 3.03 (1H, br.d, J=5.8 Hz, OH), 3,20~4.10 (5H, m), 4,16 (2H, q, J=7 Hz), 4.71 (1H, dd, J=3.6, 4.4 Hz); HR-MS m/z 2171067 (M⁴-H). Calc for C1₀H1705; 217,1076. Since the separation of the diastereomeric mixture of (2R,45,5R)-12b and 12c by HPLC was difficult, the optical purity of (2R,45,5R)-12a was determined as follows. (a) Preparation of an equilibrium mixture of (2R,45,5R)-12a. A soln of (2R,45,5R)-12a (9.5 mg, 0.043 mmol) and a catalytic amount of p-TeOH:H2O in 99 % EtOH (0,5 ml) was stirred for a week at room temp and then diluted with Bt₂O, washed with sat NaHO3 aq soln and brine, dried (MgSO4) and concentrated in vacuo to give (2R,45,5R)-12a (10.9 mg). (b) HPLC analysis of the MTPA ester, (2R,54,5R)-12b, prepared from (2R,45,5R)-12a and (6)-MTPA-Cl²⁷: [under the same condition as described for that of (25,45,5R)-12b, R 3.3 min [70.8 %, (2R,45,5R)-12a), 58.0 min [28.3 %, (25,45,5R)-12b]. (25,45,5R)-12b was shown to be a single diastereomer. Therefore the optical purity of (2R,45,5R)-12a was determined to be 100 % e.e.

<u>Ethyl</u> (45,5R)-4-Acetoxy-2-ethoxy-5-tetrahydrogyrancarboxylate 12d. (2 \overline{RS} ,4 \overline{S} ,5 \overline{R})-12a (4.7 g, 22 mmol) was treated with Ac₂O (6.2 ml, 65.7 mmol) and 4-(N,N-dimethylamino)pyridine (DNAP, 0.25 g, 2.0 mmol) in C₅H₅N (20 ml) in a usual manner followed by chromatographic purification over SiO₂ (70 g, n-hexane-EtOAc (9:1)) to give 12d (6.0 g, quantitative). A small amount of (2 \overline{S} ,4 \overline{S} ,5 \overline{R})-12d was obtained from the later eluted fractions: $n_{\rm p}^{\rm f0}$ 1.4419; (α) $_{\rm p}^{\rm f7}$ +32.7° (c=1.07); vmax 1740 (8), 1240 (8), 1125 (a), 1025 (s) cm⁻¹; & (CCl₄) 1.19 (3H, t, J-7 Hz), 1.23 (3H, t, J-7 Hz), 1.97 (3H, e), 1.70-2.20 (1H, m), 2.20-2.49 (1H, m), 2.66 (1H, ddd, J=3.2, 4.8, 11 Hz), 3.00-4.20 (4H, m), 4.12 (2H, q, J-7 Hz), 4.67 (1H, dd, J=2, 4 Hz), 5.21 (1H, ddd, J=3.2, 3.2, 12.2). (Found: C, 55.30; H, 7.73. Calc for Cl₂H_{2O}O₆: C, 55.37; H, 7.75 b).

(48,5R)-2-Ethoxy-4-hydroxy-5-tetrahydropyranmethanol 13a. 12d (58 g, 22.3 mmol) was reduced with LAH (1.75 g, 46.1 mmol) in Et₂O (100 ml) in a usual manner to give crude 13a (3.8 g, 97 %), ymax 3420 (s), 1120 (s), 1030 (s) cm⁻¹; & (CC14) 1.25 (3H, t, J=7 Hz), 1.45 (2H, s, OHx2), 1.55~2.40 (3H, m), 2.80~4.60 (7H, m), 4.95 (1H, br.dd, J=3, 3 Hz); MS m/z 176 (M⁺). This was employed in the next step without further purification.

 $\frac{(2R,4S,5S)-}{(2R,4S,5S)-2-ethoxy-4-hydroxy-5-tetrahydropyranmethanol}{(2R,4S,5S)-13c} (13c, 13a (44 mg, 0.25 mmol) was treated with benzoyl chloride (0.12 ml, d 1.211, 1.0 mmol) in C_{5H5}M (0.7 ml) in a usual manner followed by chromatographic purification over SiO₂ [2 g, n-hexane-EtOAc (40:1~30:1)] to give (2R,4S,5S)-13c (91 mg, 95 %). HELC (n-hexane-THF (20:1), 1.1 ml/min] Rt 31.9 min (37 %), 52.7 min (63 %). Further purification of this by prep TLC [n-hexane-THF (20:1), 1.1 ml/min] Rt 31.9 min (37 %), 52.7 min (63 %). Further purification of this by prep TLC [n-hexane-Et₂O (5:1, x 8)] gave (2R,4S,5S)-13c (19 mg, 21 %) as a less polar isomer and (2S,4S,5S)-13c (47 mg, 52 %) as a more polar isomer. (2R,4S,5S)-13c (19 mg, 21 %) as a less polar isomer and (2S,4S,5S)-13c (47 mg, 52 %) as a more polar isomer. (2R,4S,5S)-13c (19 mg, 21 %) as a less polar isomer and (2S,4S,5S)-13c (47 mg, 52 %) as a more polar isomer. (2R,4S,5S)-13c (19 mg, 21 %) as a less polar isomer and (2S,4S,5S)-13c (47 mg, 52 %) as a more polar isomer. (2R,4S,5S)-13c (19 mg, 21 %) as a less polar isomer and (2S,4S,5S)-13c (47 mg, 52 %) as a more polar isomer. (2R,4S,5S)-13c (19 mg, 21 %) as a less polar isomer and (2S,4S,5S)-13c (47 mg, 52 %) as a more polar isomer. (2R,4S,5S)-13c (19 mg, 21 %) as a less polar isomer and (2S,4S,5S)-13c (17 mg, 52 %) as a more polar isomer. (2R,4S,5S)-13c (19 mg, 21 %) as a less polar isomer and (2S,4S,5S)-13c (10 m, m) +11 (237), 0 (227), -2.8 (220); wax 3090 (w), 3040 (w), 1725 (vs), 1605 (m), 1590 (m), 1270 (vs), 1110 (vs), 715 (vs), 690 (m) cm⁻¹, 6 (100 MHz) 1.26 (3H, t, J=7.3 Hz), 1.97 (1H, dd, J=4.4, 4.4, 13 Hz), 2.17 (1H, ddd, J=3.3, 7.3, 13.0), 2.40-2.77 (1H, m), 3.50 (1H, dg, J=7.3, 10 Hz), 3.84 (1H, dd, J=4.1, 4.4, 13 Hz), 2.17 (1H, ddd, J=4.4, 12 Hz), 4.52 (2H, d, J=7.3 Hz), MS m/z 384 (M⁺), 369, 339, 262, 217, 140, 123, 122, 105, 77. (2S,4S,5S)-13c (TL [n-hexane-EtOAc (2:1)] Rf 0.667 CD (c=1.0 x 10⁻², EtOH, t=25^{5}): [\Delta E (\lambda, nm)] +5.8 (238), 0 (222), -1.0 (220), wax 3080 (w), 3050 (w$

 $\frac{(45,5R)-2-Ethoxy-4-hydroxy-5-tetrahydrogyranmethanol dibenzyl ether 13b. Under Ar 13a (3,7 g, 21 mmol) was treated with NaH (3,5 g, 60 % in mineral oil, 87,5 mmol), <u>n-Bught I</u> (0,38 g, 1.0 mmol)³⁰ and PhCH₂Br (10,9 g, 63,8 mmol) in THF (80 ml) with refluxing for 1 h followed by chromatographic purification over Sio₂ [70 g, <u>n-bexane-EtOAc</u> (50:1-10:1)] to give 13c (6.0 g, 80 %), <math>n_0^{77}$ 1.5267; $(a)_0^{21}^{-1}$ -16.3° (c=0.71); vmax 3110 (w), 3080 (m), 3050 (m), 1500 (m), 1100 (s), 740 (s), 700 (s) cm⁻¹ 5 (cCl₄) 1.15 (3H, t, J=7 Hz), 1.50-1.87 (2H, m), 1.87-2.33 (1H, m), 3.00-4.21 (7H, m), 4.39 (4H, br.s), 4.49~4.82 (1H, m), 7,17 (10H, br.s); MS <u>m/z</u> 356 (M⁺), 341, 310, 241, 219, 203, 202, 91.

 $\frac{(48,5R)-4-Benzyloxy-5-benzyloxymethyl-2-tetrahydrogyranyltriphenylphosphonium tetrafluoroborate 14. A soln of 13b (1.13 g, 3.17 mmol) and Ph₃PH⁺BF₄⁻ (1.26 g, 3.61 mmol)³¹ in MeCN (20 ml) was stirred and heated under reflux for 1 h under Ar. The mixture was concentrated <u>in vacuo</u>, and the residue was washed with 8E₂O (10 ml x 13). The residue was then suspended in C₆H₆ (20 ml), and the mixture was concentrated <u>in vacuo</u> to remove H₂O azeotropically. This process was repeated four times. The residue was heated at ~60° (bath temp) <u>in vacuo</u> overnight to dryness, giving crude 14 (2.20 g, quantitative). This was employed in the next step without further purification.$

Ethyl $(25,1^{15})-2-(1^{1}-hydroxyethyl)pent-4-encate (25,1^{15})-17a$ According to the reported procedure, ³³ (5)-16 [b.p. 70~75°/15 Torr; $(a)g^{1} + 42.6^{\circ}$ (c=1.04); 96~98 % e.e.; 26.4 g, 0.20 mol] gave $(25,1^{15})-17a$ (29.3 g, 85.2 %), b.p. 95~103°/16 Torr; $[a]g^{2} + 14.8^{\circ}$ (c=1.55); $[11t_{*}^{33} [a]g^{2} + 14.5^{\circ}$ (c=0.37, CHCl₃); ca. 97 % e.e.].

Ethyl $(2RS,1^{1}RS)-2-(1^{+}hydroxyethyl)pent-4-enoate (2RS,1^{1}RS)-17a.$ Ethyl 2-acetylpent-4-enoate [5,4 g, prepared from ethyl acetoacetate 15, allyl bromide and $K_{2}CO_{3}$ in refluxing acetone-DMP (4:1); contaminated with dialkylated product (purity 60 %)] was reduced with NaBH₄ (0.49 g, 10.3 mmol) in 99 % EtOH (30 ml) in a usual manner. Chromatographic purification (SiO₂ (140 g), <u>n</u>-hexane-Et₂O (12:1~10:1)] gave ethyl 2-allyl-2-(1'-hydroxyethyl)pent-4-enoate (2.1 g, 26 % from 15) and (±)-17a (2.1 g, bp, 61~65°/3 Torr; 32 % from 15).

Ethyl (25,1'5)-2-(1'-tetrahydropyranyloxyethyl)pent-4-enoate 17b. (25,1'5)-17a (29,1 g, 169 mmol) was treated with dihydropyran (23,1 g, 274 mmol) and pyridinium p-toluenesulfonate (PPTS, 5.5 g, 21.9 mmol) an $C_{12}C_{12}$ (150 ml) followed by a usual workup $C_{10}C_{11}$ to give 17b (43,7 g, quantitative), bp 92°/0.2 Torr, n_{2}^{4} 1.4476; $(\alpha)_{3}^{2}$ +25.3° (c=1.07); vmax 3110 (m), 1740 (s), 1650 (m), 1185 (s), 1125 (s), 1035 (s), 1025 (s), 990 (s), 915 (s) cm⁻¹; 6 (CCl₄) 1.11 (3H, d, J=7 Hz), 1.24 (3H, t, J=7 Hz), 1.35~2.00 (6H, m), 2.36 (2H, br.d, J=5 Hz), 1.97~2.65 (1H, m), 3.10~4.40 (3H, m), 4.08 and 4.10 (total 2H, each q, J=7 Hz), 4.45~4.78 (1H, br.d, J=8 Hz), 4.78~5.25 (2H, m), 5.25~6.30 (1H, m). (Found: C, 65.29; H, 9.44. Calc for $C_{14}H_2A_{24}$; C, 65.69; H, 9.44 %).

 $\frac{(2R,3S)-2-Allylbutane-1,3-diol}{1} = \frac{3-HP}{2} \frac{ether}{18a} = 17b (43.5 g, 169.7 mmol) was reduced with LAH (7.5 g, 198 mmol) in Et₂O (450 ml) in a usual mannerCf.10 to give 18a (36.3 g, 99.7 %), bp. 100-101°/0.2 Torr, ng⁵ 1.4634; [a]g⁴ +16.2° (c=1.10); vmax 3470 (s), 3100 (m), 1645 (m), 1025 (s), 995 (s), 910 (m) cm⁻¹; 6 (CCl₄) 1.15 and 1.25 (total 3H, each d, J=7 Hz), 1.25-1.95 (7H, m), 2.12 (2H, dd, J=7, 7 Hz), 2.45 and 2.57 (total 1H, each dd, J=5, 5; 7 Hz, OH), 3.12~4.30 (5H, m), 4.40~4.75 (1H, br.s), 4.75~5.35 (2H, m), 5.35~6.30 (1H, m). (Found: C, 66.84; H, 10.56. Calc for C₁₂H₂₂O₃: C, 67.25; H, m) (5H, m) (5$

10.35 %).

 $\frac{(2R,35)-2-A11y1butane-1,3-diol}{2} \frac{1-benzy1}{2} \frac{3-THP}{2} \frac{1}{2} \frac{1}{1} \frac{1}{2} \frac{1}{2} \frac{1}{1} \frac{1}{2} \frac{1}$

 $\begin{array}{l} 19.9293 \text{ (2R,35)-2-Allylbutane-1,3-diol} 1-benzy1 \text{ ether } 18c. (2R,35)-18b (59.0 g, ~167 mmol) was treated with p-TsOH+H_2O (1.15 g, 6.0 mmol) in MeOH (500 ml). After a usual workup, Cf.11 chromatographic purification [SiO_2 (700 g), n-hexane-EtOAc (10:1-8:1)] gave 18c (38.4 g, quantitative). Analytical sample: b,p. 108°/0.17 Torr, <math>n_{2}^{64}$ 1.5056; $[\alpha]_{2}^{23}$ -1.50° (c=1.47); vmax 3520 (s), 3100 (m), 3060 (m), 3010 (s), 1645 (m), 1500 (m), 1010 (s), 1000 (s), 915 (s), 740 (s), 700 (s) cm^{-1}; 6 (CCl_4) 1.12 (3H, d, J=7 Hz), 1.36~1.85 (1H, m), 2.12 (2H, br.dd, J=6, 7 Hz), 2.55~2.75 (1H, m, OH), 3.46 (2H, dd, J=3, 5 Hz), 3.71 (1H, dq, J=6, 6 Hz), 4.41 (2H, s), 4.72~5,20 (2H, m), 5.35~6.15 (1H, m), 7.27 (5H, br.s); Capillary GLC (Column, P2G 20M, 50 m x 0.25 mm at 170°; Carrier gas, N₂, 1.1 kg/cm²) Rt 38.8 min (96.7 %), 39.9 min (3.3 %). (Pound: C, 75.96; H, 9.01. Calc for C1_4H2_0O_2: C, 76.32; H, 9.15 %). \\ \end{array}

 $\frac{(2R,3S)-2-Allylbutame-1,3-diol}{(2R,3S)-2-Allylbutame-1,3-diol} \frac{1-benzyl}{1-benzyl} ether \frac{3-(3^*,5^*-dinitro)benzoate}{(2R,3S)-2-Allylbutame-1,3-diol} \frac{1-benzyl}{1-benzyl} ether \frac{3-(3^*,5^*-dinitro)benzoate}{(2R,3S)-2-Allylbutame-1,3-diol} \frac{1-benzyl}{1-benzyl} ether \frac{3-(3^*,5^*-dinitro)benzoate}{(2R,3S)-2-Allylbutame-1,3-diol} \frac{1-benzyl}{1-benzyl} ether \frac{3-(3^*,5^*-dinitro)benzoate}{(2R,3S)-2-Allylbutame-1,3-diol} \frac{1-benzyl}{1-benzyl} ether \frac{3-(3^*,5^*-dinitro)benzoate}{(3R,5)} \frac{18c}{(3R,9)} (140 ml) in a usual manner cf.11 to give crude 18d (60.7 g, 92 %). This was recrystallized four times from n-bexane-Et_2O (3:1) followed by another recrystallization from n-bexane-Et_2O (100:1) to give pure 18d (44.6 g, 73.5 % recovery) as yellowish needles, m.p. <math>61-62^\circ$; $(a)_1^2 + 42.1^\circ (c=2.50)$; wmax 3150 (w), 3120 (m), 1720 (s), 1655 (w), 1630 (m), 1600 (w), 1550 (s), 1500 (w), 1350(s), 1280 (s), 1180 (s), 925 (s), 880 (m), 760 (m), 735 (s), 730 (s), 720 (s), 705 (m) cm^{-1}; \delta (CDC1_3) 1.44 (3H, d, J=6 Hz), 1.90-2.50 (1H, m), 2.20 (2H, dd, J=6, 6 Hz), 3.55 (2H, br.d, J=5 Hz), 4.45 (2H, s), 4.86-5.30 (2H, m), 5.30-6.20 (2H, m), 5.40-6.25 (1H, m), 5.55 (1H, dq, J=6, 6 Hz), 7.25 (5H, br.s), 9.10 (2H, d, J=2 Hz), 9.21 (1H, t, J=2 Hz). HPLC [n-bexane-THF (100:1), 1.3 ml/min] Rt 44.2 min (99.9 %). Its diastereomer eluted at Rt 42.3 min. (Found: C, 61.17; H, 5.39; N, 6.75. Calc for $C_{21H22}O_7N_2$: C, 60.266 H, 5.35; N, 6.76 %).

Diastereomerically and enantiomerically pure (2R,3S)-2-Allylbutane-1,3-diol 1-benzyl ether 18c. (2R,3S)-18d (44.6 g, 108 mmol) was hydrolyzed with N KOH aq soln (150 ml, 150 mmol) in THF-99 % EtOH (1:1, 360 ml)^{cf.11} to give 18c (23.5 g, 99 %), b.p. 108-109°/0.16 Torr; n_{c}^{c5} 1.5045; $[\alpha]_{c}^{c5}$ -1.96° (c=2.08); its IR and NMR spectra were almost identical with those described above. Capillary GLC (under the same condition as described above) Rt 18.4 min (100 %). Its diastereomer eluted at Rt 19.7 min. (Found: C, 76.28; H, 9.17. Calc for $C_{14}H_{20}O_{2}$: C, 76.32; H, 9.15 %). According to the reported procendure,²⁷ (2R,3S)-18e was prepared from (2R,3S)-18e and (S)-MTPA chloride (MTPA-Cl). HPLC [n-hexane-THF (100:1), 1.3 ml/min] Rt 18.0 min [single peak, (2R,3S)-18e]. Its diastereomer (2S,3R)-18e prepared from (2R,3S)-17a eluted at Rt 19.2 min.

(2R,35)-2-Allylbutane-1,3-diol 1-benzyl ether 3-tosylate 18f. (2R,35)-18c (12 g, 54,5 mmol) was treated with p-TSCl (25 g, 130 mmol) and DMAP (1.46 g, 12 mmol) in CeHeN (30 ml) in a usual mannerCf.10,11 to give 18f (20.5 g, quantitative). This was employed in the next step without further purification; wmax 3100 (w), 3060 (w), 3020 (m), 1645 (m), 1605 (m), 1500 (w), 1360 (m), 1180 (s), 1100 (s), 910 (s), 815 (m), 780 (m), 735 (m), 700 (m) cm⁻¹; & (CCl₄) 1.21 (3H, d, J=7 Hz), 1.61~2.28 (3H, m), 2.38 (3H, s), 3.29 (2H, d, J=7 Hz), 4.30 (2H, s), 4.50~5.20 (3H, m), 5.25~6.10 (1H, m), 7.25 (2H, d, J=8 Hz), 7.26 (5H, br.s), 7.75 (2H, d, J=8 Hz).

(R)-5-Benzyloxy-4-ethyl-1-pentere 19a. Super Hydride[®] (LiBEt₃H, 1 M in THF, 64 ml, 64 mmol) was added dropwise to a stirred soln of 18f (12.1 g, 32 mmol) in THF (36 ml) below -5° under Ar. The mixture was stirred and heated under reflux for 20 min. After cooling, to the mixture ware added H₂O (5 ml), 3 N NaOH aq soln (30 ml) and 35 % H₂O₂ aq soln (27 ml) successively below 50° with ice-cooling. The mixture was stirred for 30 min at room temp, then concentrated in vacuo to remove THF and EtOH. The residue was extracted with Et₂O (50 ml x 5). The extract was washed with H₂O and brine, dried (MgSO₄) and concentrated in vacuo. The residue (7.0 g) was chromatographed over SiO₂ (70 g n-pentane-Et₂O (20:1)] to give 19a (6.5 g, quantitative). Analytical sample: b.p. 90°/0.3 Torr, n_2^{5} 1.4892; $[\alpha]_2^{6}$ +4.62° (c=1.36), vmax 3100 (w), 3050 (w), 3000 (s), 1645 (m), 1500 (w), 1100 (s), 995 (m), 910 (s), 735 (s), 700 (s) cm⁻¹, 6 (CCl₄) 0.86 (3H, t, J=6 Hz), 1.08~1.85 (3H, m), 2.11 (2H, dd, J=6, 6 Hz), 3.30 (2H, d, J=5 Hz), 4.41 (2H, s), 4.75~5.22 (2H, m), 5.40~6.20 (1H, m), 7.30 (5H, br.s). (Found: C, 82.34; H, 9.70. Calc for C₁₄H₂₀O: C, 82.30; H, 9.87 %).

 $\frac{(R)-2-Ethyl-4-penten-1-ol}{(R)-19b}$ To a soln of (R-19a (6.5 g, 32 mmol) in THF (16 ml) and liq. NH₃ (-80 ml) was added finely cut Na (1.66 g, 72.2 mmol) portionwise with dry ice-acetone cooling and stirring. The mixture was refluxed for 10 min with stirring. To the dry ice-acetone cooled mixture was added solid NH₄Cl to discharge its deep blue color. NH₃ was evaporated from the mixture with stirring at room temp. The residue was diluted with Et₂O and filtered through a pad of Celite. The solid was washed with Et₂O. The combined filtrate and washings were washed with brine, dried (MgSO₄) and concentrated in vacuo to give crude (R)-19b (7.3 g, contaminated with PhNe). Analytical sample: b,p. 92-93°/53 Torr; n₁^B 1.4394; $[a]_{2}^{19}-0.75^{\circ}$ (c=1.56); vmax 3350 (s), 3070 (m), 1655 (s), 1040 (s), 990 (s), 910 (s) cm⁻¹; δ (CC1₄) 0.88 (3H, t, J=6 Hz), 1.05~1.60 (3H, m), 2.06 (2H, dd, J=6, 6 Hz), 2.86 (1H, br.t, J=5 Hz, OH), 2.43 (2H, br.dd, J=5, 5 Hz), 4.65~5.25 (2H, m), 5.30~6.20 (1H, m). (Found: C, 73.44; H, 12.21. Calc for C7H₁₄O: C, 73.64; H, 12.36 %).

 $\frac{(R)-4-Ethyl-5-tetrahydropyranyloxy-1-pentene}{(R)-4-Ethyl-5-tetrahydropyranyloxy-1-pentene} 19c. Crude 19b (7.3 g, ~32 mmol) was treated with dihydropyran (9.0 g, 107 mmol) and PPTS (4.0 g, 16 mmol) in CH₂Cl₂ (45 ml) as described above followed by chromatographic purification [SiO₂ (70 g), n-pentane-Et₂O (30:1)) then distillation in the presence of K₂O₃ to give 19c (6.1 g, quantitative from pure (2R, 3S)-18c], b,p. 77°/3 Torr; ng² 1.4481; (a)g²¹ +3.00° (c=1.07), vmax 3080 (m), 1640 (m), 1120 (s), 1030 (s), 990 (s), 910 (s) cm⁻¹; 6 (CCl₄) 0.92 (3H, t, J=6 Hz), 1.15-2.00 (9H, m), 2.12 (2H, dd, J=6, 6 Hz), 2.98~4.10 (4H, m), 4.40~4.65 (1H, br.s), 4.75~5.30 (2H, m), 5.40~6.20 (1H, m). (Found: C, 72.62; H, 11.14. Calc for Cl₂H₂2O₂: C, 72.62; H, 11.18 %).$

(R)-3-(Tetrahydropyranyloxymethyl)pentanal 20. To a stirred two-phase mixture of 19c (1.5 g, 7.9 mmol) in Et₂O-H₂O (1.1, 40 ml) were added NaIO₄ (5.6 g, 26.2 mmol) and 5 & OBO₄ soln in THE (2.4 ml, 0.47 mmol) at room temp. After stirring vigorously for 2 h at room temp, an additional amount of NaIO₄ (3 g, 14 mmol) was added to the mixture. The vigorous stirring was further continued for 5.5 h at room temp. The precipitate was filtered of through a pad of Celite and washed with Et₂O. The organic layer was separated from the combined filtrate and washings, and the aq layer was extracted with Et₂O. The combined Et₂O soln was washed with Na₂S₂O₃ aq soln, sat NaHCO₃ aq soln and brine, dried (MgSO₄) and concentrated in vacuo. The residue was distilled under Ar to give 20 (1.02 g, 65 %), b.p. 94°/2.0 Torr; n_{β}^{2} 1.4529; $[\alpha]_{\beta}^{2}$ +25.2° (c=1.07); vmax 2740 (m), 1725 (s), 1125 (s), 1035 (s) cm⁻¹; 6 (CCl₄) 0.92 (3H, t, J=7 Hz), 1.15~2.15 (9H, m), 2.15~2.50 (2H, m), 2.30~3.96 (4H, m), 4.30~4.60 (1H, br.s), 9.74 and 9.79 (total 1H, each d, J=2 Hz). (Found: C, 66.07; H, 10.06. Calc for C₁₁H₂O₂: C, 65.97; H, 10.07 %).

 $(3^{1}R,45,5R)-4$ -Benzyloxy-5-benzyloxymethyl-2- $[(3^{4}-tetrahydropyranyloxymethyl)pentylidene)tetrahydropyran 22. To a stirred$ and cooled soln of crude 14 (2.20 g, ~3.17 mmol) and HMPA (5.5 ml) in THF (29 ml) was added dropwise <u>n</u>-BuLi (1.72 M in <u>n</u> $hexane, 2.1 ml, 3.61 mmol) at <math>-67^{\circ}-63^{\circ}$ under Ar. After stirring for 30 min at -67° , to this deep red ylide soln was added dropwise a soln of 20 (0.83 g, 4.12 mmol) in THF (6.5 ml) at $-67^{\circ}-63^{\circ}$ with stirring. The temp was raised gradually to room temp over 3 h 50 min and the stirring was continued for 6 h 15 min at room temp. To the mixture was added sat NAHCO₃ aq soln (18 ml) at $-10^{\circ}-0^{\circ}$ with stirring. The mixture was concentrated <u>in vacuo</u> to remove THF. The residue was extracted with $8t_2O$ (20 ml x 4). The extract was washed with sat NAHCO₃ ag soln and brine, dried (K₂CO₃) and concentrated <u>in vacuo</u> to give crude 22 (2.94 g). According to the procedure described above, 3.85 g (10.8 mmol) of 13b was converted to 9.0 g of crude 22. This was employed in the next step without further purification.

 $\frac{(3R,4S,6R,9R)-and}{(3S,4R,6S,9R)-4-Benzyloxy-3-benzyloxymethyl-9-ethyl-1,7-dioxaspiro[5.5]undecame 1b + 23b and (3R,4S,6R,9R)-, (3S,4S,6R,9R)-and (3R,4R,6S,9R)-3-benzyloxymethyl-9-ethyl-4-hydroxy-1,7-dioxaspiro[5.5]undecame 1c + 2c + 24c, Crude 22 (9,0 g) was mixed with conc HC1-H₂O-THF (1:5:0, 90 ml) with ice-salt cooling. After sturring for 15 h at room temp, the mixture was neutralized by the addition of sat NaHCO₃ ag soln with sturring and ice-cooling, and concentrated in vacuo to remove THF. The resultue was extracted with Et₂O. The extract was washed with sat NaHCO₃ ag soln and brine, dried (Mg90₄) and concentrated in vacuo. The residue (7.8 g) was chromatographed over SiO₂ [210 g, n-hexane-EtOAc (80:1-10:1)] to give a mixture (2.04 g, 46 % from 13b) of 1b and 23b, ng^A 1.5274; [3]g² - 105° (c=0.51); wmax 3100 (w), 3080 (w), 3050 (m), 1500 (m), 1105 (vs), 1080 (vs), 1005 (s), 740 (s), 700 (s) cm⁻¹; 6 (100 MHz) 0.88 and 0.90 (total 3H, each t, J=7.5 Hz), 1.00-1.80 (BH, m), 1.87 (1H, dd, J=5.2, 13.3 Hz), 2.10-2.43 (1H, m), 3.14 (-0.7H, dd, J=9.9, 9.9 Hz), 3.27-3.63 (-2.3H, m), 3.73 (2H, d, J=6.9 Hz), 4.95 and 4.97 (total 1H, each dd, J=3.7, 11.7; 4.0, 12.0 Hz), 4.06 (1H, ddd, J=5.1, 5.1, 11.0 Hz), 4.43 (1H, d, J=11.7 Hz), 4.47 (1H, d, J=11.7 Hz), 4.59 (1H, d, J=11 Hz), 4.62 (1H, d, J=11.7 Hz), 7.28 (-6H, br.s), 7.33 (-4H, br.s). (Found: C, 75.63; H, 8.38. Calc for C₂₆H₃₄O₄: C, 76.06; H, 8.34 %). Further elution [n-bexane-EtOAc (10:1-3:1)] gave a mixture (1,21 g, 35 % from 13b) of 1b and 24c, vmax 3430 (s), 3100 (w), 3070 (w), 2004 (w), 1500 (w), 1075 (s), 740 (m), 700 (m) cm⁻¹; 6 (CC14) 0.88 (3H, t, J=6.4 Hz), 1.05-1.50 (10H, m), 2.90-4.30 (8H, m), 4.49 (2H, br.s), 7.32 (5H, br.s); MS m/z 320 (M⁺), 302, 129 127, 126, 125, 91 (base peak).$

(38,45,68,98)-, (35,45,68,98)- and (38,48,65,98)-4-[3-Benzyloxymethyl-9-ethyl-1,7-dioxaspiro(5.5]undecyl] benzoate 1f, 2f and 24f. A mixture of 1c, 2c and 24c (25 mg, Q.078 mmol) was treated with benzoyl chloride (Q.012 ml, d 1.211, 0.10 mmol) in C5H5N (0.5 ml) in a usual manner followed by purification with prep TLC (n-hexane-EtOAc (7:2)) to afford 28 mg (82 %) of 1f, 2f and 24f, vmax 3110 (w), 3080 (w), 3050 (w), 1725 (s), 1605 (w), 1590 (w), 1495 (w), 1270 (s), 1100 (s), 740 (m), 715 (s) cm⁻¹, 6 (100 MHz) 0.89, 0.91 and 0.96 (total 3H, each t, J=7.3 Hz), 1.00~2.00 (7H, m), 1.97 (1H, dd, J=5.1, 13.1 Hz), 2.10~2.53 (1H, m), 3.41 (~0.33H, dd, J=9.5, 9.5 Hz, 1f), 3.52 (~0.33H, dd, J=13.1, 13.1 Hz, 2f), 3.56 (~0.33H, dd, J=3.7, 13.1 Hz, 2f), 3.61 (~0.33H, br.d, J=11.7 Hz, 24f), 3.55~3.65 (~0.33H, m, 1f), 3.60~3.80 (2H, m), 3.75 (~0.33H, dd, J=5.1, 11.7 Hz, 24f), 3.73~3.82 (~0.66H, m, 1f), 3.87 (~0.67H, dd, J=11.7, 11.7 Hz, 2f + 24f), 3.96 (~0.67H, dd, J=5.1, 11.7 Hz, 2f + 24f), 4.42 (~0.67H, g), 4.45 (~0.67H, g), 4.53 (~0.66H, s), 5.36 (~0.67H, dd, J=5.1, 11. 11 Hz, 2f + 24f), 5.62 (~0.33H, dd, J=5.5, 5.5, 11.7 Hz, 1f), 7.26 and 7.29 (total 5H, each s), 7.36~7.75 (3H, m), 7.83~8.20 (2H, m); MS m/z 424 (M⁺), 423, 321, 312, 302, 129, 127, 126, 123, 122, 105, 91, 77.

 $\frac{(3R,4S,6R,9R)-}{(3R,4S,6R,9R)-} and \frac{(3S,4R,6S,9R)-9-Ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro[5,5]undecame 1a and 23a. A mixture of 1b and 23b (2,35 g, 5,72 mmol) was treated with Na (1.41 g, 61,3 mmol) in THF (6 ml) and lig, NH₃ (-80 ml) in the usual manner to give crude 1a and 23a (0,99 g, 75 %), vmax 3370 (s), 1070 (s) cm⁻¹, & (CDCl₃) 0,88 (3H, distorted t, J=7 Hz), 2,35 (2H, s, OHz2), 1,1~2.4 (10H, m), 2,73~4.10 (6H, m), 4.10~4.65 (1H, m). This was employed in the next step without further purification.$

 $\frac{(3S, 4S, 6S, 9R)-,}{(3R, 4R, 6S, 9R)-,} \frac{(3R, 4R, 6S, 9R)-}{(3R, 4R, 6S, 9R)-} and \frac{(3S, 4S, 6S, 9R)-3-[9-Ethyl-4-hydroxy-1,7-dioxaspiro[5.5]undecyl]methyl}{(3'5'-dinitro)benzoate} le [talaromycin A 12-(3',5'-dinitro)benzoate], 23e, 25e and 26e. A mixture of crude la and 23a (0.99 g, 4.3 mmol) was treated with DMB-Cl (1.10 g, 4.75 mmol) and DMAP (16 mg, 0.13 mmol) in C_{5H5}N (18 ml) with ice-salt cooling. The stirring was continued for 3 days at room temp. After a usual workup,¹¹ the residue (1.8 g) was chromatographed over SiO₂ (40 g). Firstly eluted fractions [C_{6H6}-EtOAc (80:1-60:1)] gave 487 mg (18 %) of 1d and 23d as crystals, TLC [n-hexane-EtOAc (3:1)] Rf 0.53, vmax 3120 (m), 1730 (s), 1630 (m), 1600 (w), 1550 (s), 1350 (s), 1280 (s), 1165 (s), 760 (s), 725 (s) cm⁻¹, 6 (CCCl₃) 0.70~1.70 (3H, m), 1.10~2.15 (8H, m), 1.93~2.35 (1H, m), 2.35~2.90 (1H, m), 3.25~4.70 (4H, m), 4.77 (2H, brzl, J=6 Hz), 5.50~6.10 (1H, m), 9.00~9.37 (6H, m); MS m/z 406 (M⁺-DNBOH).$

Secondly eluted fractions (40:1~10:1) gave 25e and 26e (254 mg, 14 $\overline{9}$, TLC [n-hexane-EtOAc (3:1)] Rf 0.36; vmax 3530 (s), 3120 (s), 1735 (s), 1630 (m), 1600 (w), 1550 (s), 1350 (s), 1280 (s), 1080 (s), 775 (m), 760 (m), 730 (s), 725 (s) cm⁻¹, δ (CDCl₃) 0.89 (3H, br.t, J~5 Hz), 1.10~2.20 (8H, m), 2.00~2.70 (2H, m), 3.10~4.37 (5H, m), 4.44 and 4.50 (total 2H, each br.d, J=7.0 Hz), 4.65~5.10 (1H, m), 9.23 (2H, d, J=2 Hz), 9.30 (1H, t, J=2 Hz), NS m/z 424 (M⁺). 36 mg of this was further purified by prep TLC [n-hexane-EtOAc (3:1, x 2)] to afford 26e (16 mg, 44 $\overline{9}$) as a less polar isomer and 25e (5 mg, 14 $\overline{9}$) as a more polar isomer. 26e: TLC [n-hexane-EtOAc (3:1 x 3)] Rf 0.62; vmax 3530 (s), 3120 (s), 1735 (s), 1630 (s), 1600 (m), 1550 (s), 1350 (s), 1280 (s), 1170 (s), 770 (s), 730 (s), 720 (s) cm⁻¹, δ (400 MHz) 0.93 (3H, t, J=7.3 Hz), 1.37~1.64 (5H, m), 1.68 (1H, dd, J=3, 14 Hz), 1.70 (1H, ddd, J=5.0, 13.5, 13.5, 12.), 2.00 (1H, dddd, J=4.5, 4.5, 13.5, 13.5, Hz), 2.03 (1H, dd, J=3, 14 Hz), 2.23~2.33 (1H, m), 3.52 (2H, br.d, J=11 Hz), 3.74 (1H, dd, J=5.11 Hz), 3.81 (1H, dd, J=11, 11 Hz), 3.86 (1H, dd, J=2.8, 11 Hz), 3.92 (1H, d, J=10 Hz, OH), 4.07~4.13 (1H, m), 4.40 (1H, dd, J=7.5, 11 Hz), 4.55 (1H, dd, J=7.0, 11 Hz), 9.14 (2H, d, J=2 Hz), 9.23 (1H, t, J=2 Hz). 25e: TLC [n-hexane-EtOAc (3:1, x3)] Rf 0.58; its physical and spectral data were described later.

Thirdly eluted fractions (10:1-5:1) gave 1e and 23e (730 mg, 40 %) as crystals, TLC [n-hexane-EtOAc (3:1)] Rf 0.13. This mixture was further purified by fractional recrystallization, and the mother liquor was also purified by prep TLC [n-hexane-EtOAc (3:1, x 5)] to afford 23e (123 mg, 17 %) as a less polar isomer and 1e (509 mg, 70 %) as a more polar isomer. 23e: TLC [n-hexane-EtOAc (3:1, x4)] Rf 0.40; m.p. 171.5-173.0° (needles); [α] $_{10}^{16}$ +136° (c=0.63); vmax 3460 (s), 3120 (w), 2990 (m), 1750 (s), 1630 (m), 1610 (w), 1550 (s), 1350 (vs), 1280 (s), 1170 (s), 735 (m), 730 (m), 725 (m) cm⁻¹, 6 (400 MHz) 0.93 (3H, t, J=7.3 Hz), 1.19 (2H, dq, J=7.3, 5.3 Hz), 1.35-1.55 (3H, m), 1.54-1.64 (1H, m), 1.66 (1H, ddd, J=4.5, 13.0, 13.0, Hz), 1.74-1.87 (1H, br.s, OH), 1.94 (1H, dd, J=5.6, 13.0 Hz), 2.00 (1H, dddd, J=4.5, 4.5, 13.0, 13.0 Hz), 2.32-2.39 (1H, m), 3.44 (1H, ddd, J=11.5, 3.0, 3.0 Hz), 3.79 (1H, dd, J=3.0, 11.5 Hz), 3.84 (2H, seemingly d, J=2.0 Hz), 4.43 (1H, ddd, J=5.6, 5.6, 11.2 Hz), 4.62 (1H, dd, J=9.8, 11.0 Hz), 4.84 (1H, dd, J=4.0, 11.0 Hz), 9.16 (2H, d, J=2.0 Hz), 9.23 (1H, t, J=2.0 Hz). (Found: C, 54.161 H, 5.541 N, 65.0. Calc for C19H2409N2: C, 53.771 H, 5.705 N, 6.60 %). 1e: TLC [n-hexane=BtOAc (3:1, x 3)] Rf 0.34, m.p. 147-148° (fine needles); [a] $_{10}^{16}$ -113° (c=2.13); vmax 3460 (s), 3130 (m), 3100 (m), 1730 (s), 1630 (m), 1600 (w), 1550 (s), 1350 (s), 1290 (s), 1170 (s), 1080 (s), 735 (s), 725 (s) cm⁻¹, δ (400 MHz) 0.89 (3H, t, J=7.5 Hz), 1.07-1.25 (2H, m), 1.41 (1H, dddd, J=3.5, 12.0, 12.0, Hz), 1.42-1.61 (4H, m), 1.61-1.72 (1H, m), 1.75 (1H, ddd, J=3.5, 3.5, 13.0, Hz), 3.79 (1H, dd, J=5.6, 12.0 Hz), 3.83 (1H, dd, J=-2.0, Hz), 4.44 (1H, ddd, J=5.6, 1.2, Hz), 4.62 (1H, dd, J=9.8, 11.0 Hz), 4.84 (1H, dd, 1-4.0, 11.0 Hz), 9.16 (2H, d, -2.0, Hz), 4.44 (1H, ddd, J=-5.6, 5.6, 11.2 Hz), 4.62 (1H, dd, J=9.8, 11.0 Hz), 4.84 (1H, dd, 1-4.0, 11.0 Hz), 9.16 (2H, d, -2.0, Hz), 4.44 (1H, ddd, J=-5.6, 5.6, 11.2 Hz), 4.62 (1H, dd, J=9.8, 11.0 Hz), 4.84 (1H, dd, -4.0, 11.0 Hz), 9.16 (2H, d, -2.0 Hz). (Found: C, 54.11; H, 5.72; N, 6.50. Calc for C₁₉H₂₄O₉N₂: C, 53.77; H, 5.70; N, 6.60 %).

Conversion of (35,45,68,98)- and (38,48,65,98)-9-ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro[5.5]undecame bis (3',5'dinitro)benzoate 1d and 23d to 1e and 23e. A mixture of 1d and 23d (487 mg, 0,79 mmol) was treated with solid K2003 (487 mg, 0.79 mmol) in MeOH (5 ml)-THF (2.5 ml) at room temp. The crude product was treated with DNB-Cl (194 mg, 0.84 mmol) in CgH5N (4 ml) and CH2Cl2 (1 ml) followed by chromatographic purification [SiO4 (9 g), CgH6-EtOAc (25:1~6:1)] to give le and 23e [137 mg, 41 % from 1d and 23d]. This was further purified by fractional recrystallization and the mother liquor was also purified by prep TLC [n-hexane-EtOAc (3:1, x 3)] to afford 23e (26 mg, 19 %) and 1e (62 mg, 45 %).

Conversion of (1c + 2c + 24c) to 1e and (2e8 + 24e). A mixture of 1c , 2c and 24c (1.10 g, 3.44 mmol) was treated with Na (0.3 g, 13 mmol) in THP (3 ml) and liq. NH3 (~30 ml) as usual. The crude product (0.86 g) was treated with DNB-Cl (796 mg, 3.5 mmol) in C5H5N (14 ml) followed by chromatographic purification over SiO2 [20 g, C6H6-EtOAc (40:1-5:1)] to give a mixture [550 mg, 38 & from (1c + 2c + 24c)] of 1e, 2e and 24e. This was fractionally recrystallized from n-hexane-EtOAc to give pure le (93 mg, 17 %). A portion (21 mg) of the mother liquor was purified by prep TLC [n-hexane-EtOAc (4:1, x 20)] to give further amount of pure le (3 mg) and a mixture (12 mg) of 2e and 24e; vmax 3470 (s), 3130 (s), 1730 (s), 1635 (s), 1600 (m), 1550 (s), 1345 (s), 1280 (s), 1170 (s), 775 (m), 735 (s), 725 (s) cm⁻¹; 6 (100 MHz) 0.93 (3H, t, J=6.5 Hz), 1.05~2.20 (11H, m), 3.32~3.86 [total 2H; 3.50 (dd, J=12.0, 12.0, 2e), 3.57 (d, J=10.0, 24e), 3.77 (dd, J=3.0, 10.0 Hz, 24e), 3.76~3.86 (m, 2e)], 3.85 (1H, dd, J=10.0, 10.0 Hz), 3.93 (1H, dd, J=4.7, 10.0 Hz), 4.22 (1H, ddd, J=5.1, 13.0, 13.0 Hz), 4.55, 4.62, 4.63 and 4.84 (total 2H, each dd, J=5.1, 11.0; 5.1, 10.0; 4.7, 10.0; 4.4, 11.0 Hz), 9.15 and 9.18 (total 2H, each d, J=2,0 Hz), 9,26 (1H, t, J=2,0 Hz), MS: m/z 423 (M⁺-1), 406, 341, 338, 213, 129, 126,

Conversion of (2e + 23e + 24e + 25e + 26e) to (2d + 24d), 25e and (2e + 24e). A mixture of 2e, 23e, 24e, 25e and 26e (507 mg, 1.19 mmol) was treated with K2003 (0.17 g, 1.2 mmol) in MeOH (3 ml)-CHCl3 (0.5 ml) at room temp followed by chromatographic purification over SiO2 [4 g, n-hexane-EtOAc (3:1-2:1)] to give 237 mg (86 %) of an oil. This was treated with conc HCl-H_2O-THF (1:5:20, 4 ml) for isomerization.^{Cf.8} The residue (227 mg) was treated with DNB-Cl (227 mg, 0.99 mmol) in C5H5N (5 ml) followed by chromatographic purification over SiO₂ (15 g). Firstly eluted fractions [C_{6H6}-EtOAc (30:1~20:1)] gave a mixture (99 mg, 16 %) of 2d and 24d. Analytical sample [needles from <u>n</u>-hexane-EtOAc (3:1)]: m.p. 127~133°; $(\alpha)_{2}^{24}$ -2.9° (c=0.83); vmax 3120 (w), 1735 (s), 1550 (s), 1350(s), 1280 (s), 1165 (s), 735 (s), 725 (s) cm⁻¹; 6 (100 MHz) 0.91 (~1.2H, t, J=6.1 Hz, 2d), 0.95 (~1.8H, t, J=6.1 Hz, 24d), 1.05~2.13 (8H, m), 2.31 (1H, dd, J=5.8, 10.2 Hz), 2.56 (1H, dddd, J=4.7, 5.8, 11.0, 11.0 Hz), 3.27 (~0.4H, dd, J=10.6, 10.6 Hz, 2d), 3.54 (~0.6H, br.d, J=11.5 Hz, 24d), 3.63 (~0.4H, br.d, J=10.6 Hz, 2d), 3.83 (1H, dd, J=11.0, 11.0 Hz), 3.85 (~0.6H, br.d, J=11.5 Hz, 2dd), 4.03 (1H, dd, J=5.8, 11.0 Hz), 4.49 (2H, br.d, J=4.7 Hz), 5.65 (1H, ddd, J=5.1, 11.0 Hz), 9.12 and 9.14 (total 4H, each d, J=2.0 Hz), 9.25 (2H, t, J=2.0 Hz). (Found: C, 51.12; H, 4.23; N, 8.96. Calc for C26H26O14N4: C, 50.49; H, 4.24; N, 9.06 %).

Secondly eluted fractions (20:1~10:1) gave 25e (50 mg, 12 %). Analytical sample (plates from n-hexane-EtQAc): m.p. 128,8~129.2°; $[a]_{H}^{24}$ -77.7° (c=0.33); vmax 1730 (s), 1630 (w), 1545 (s), 1350 (s), 1290 (s), 1165 (m), 730(m), 725 (m) cm⁻¹; δ (400 MHz) 0.90 (3H, t, J=7.3 Hz), 1.09~1.29 (2H, m), 1.43 (1H, dddd, J=3.2, 12.5, 12.5, 12.5, Hz), 1.42~1.71 (4H, m), 1.74 (1H, dd, J=3.0, 14.5 Hz), 2.01 (1H, dd, J=3.0, 14.5 Hz), 2.23~2.33 (1H, m), 3.33 (1H, dd, J=11.0, 11.0 Hz), 3.66 (1H, ddd, J=2.0, 4.0, 11.0 Hz), 3.73 (1H, dd, J≈5.0, 11.5 Hz), 3.79 (1H, dd, J=11.5, 11.5 Hz), 3.92 (1H, d, J=10.0 Hz, OH), 4.07~4.14 (1H, m), 4.39 (1H, dd, J=7.5, 11.0 Hz), 4.56 (1H, dd, J=7.0, 11.0 Hz), 9.14 (2H, d, J=2.0 Hz), 9.23 (1H, t, J=2.0 Hz). (Found: C, 54.19; H, 5.59; N, 6.56. Calc for C19H24O9N2: C, 53.77; H, 5.70; N, 6.62 %).

Thirdly eluted fractions (10:1~5:1) gave a mixture (210 mg, 50 %) of 2e and 24e. A small amount (100 mg) was further purified by SiO₂ chromatography (3 g, CgH₆-EtOAc (20:1~8:1)) followed by prep TLC [n-hexane-EtOAc (3:1 x 3)] to give 24e as a viscous oil (61 mg, 61 %) and 2e (21 mg, 21 %) as crystals. 24e: [a] 3^4 +21.4° (c=0.51); vmax 3450 (m), 3120 (m), 1735 (s), 1635 (m), 1600 (s), 1550 (s), 1350 (s), 1280 (s), 1170 (m), 1080 (m), 735 (s), 725 (s) cm⁻¹; 6 (400 MHz) 0.92 (3H, t, J=7.3 Hz), 1.36~1.44 (2H, m), 1.45 (1H, dd, J=10.5, 12.5 Hz), 1.45~1.54 (3H, m), 1.54~1.63 (1H, m), 1.69 (1H, ddd, J=4.5, 13.5, 13.5 Hz), 1.99 (1H, dddd, J=4.5, 4.5, 13.5, 13.5 Hz), 2.03~2.09 (1H, m), 2.12 (1H, dd, J=5.0, 12.5 Hz), 3.43 (1H, br.d, J=11.5 Hz), 3.62 (1H, dd, J=11.2, 11.2 Hz), 3.78 (1H, dd, J=3.0, 11.5 Hz), 3.86 (1H, dd, J=5.0, 11.2 Hz). 3.97 (1H, ddd, J=5.0, 10.5, 10.5 Hz), 4.60 (1H, dd, J=6.0, 11.5 Hz), 4.64 (1H, dd, J=4.0, 11.5 Hz), 9.14 (2H, d, J=2.0 Hz), 9.24 (1H, t, J=2.0 Hz). (Found: C, 53.76; H, 5.75; N, 6.42. Calc for C₁₉H₂₄OgN₂: C, 53.77; H, 5.70; N, 6.60 %). 2e (fine needles from <u>m</u>-hexane-EtOAc): m.p. 104~104.2°; [a]²⁴ -29.8° (c=O.26); vmax 3460 (m), 3130 (w), 1730 (s), 1635 (m), 1550 (s), 1350 (s), 1280 (s), 1045 (s), 735 (m), 725 (m) cm⁻¹; 6 (400 MHz) 0.89 (3H, t, J≃7.5 Hz), 1.07~1.24 (2H, m), 1.41 (1H, dddd, J=3.0, 12.0, 12.0, 12.0, Hz), 1.42~1.49 (1H, m), 1.50 (1H, dd, J=11.0, 12.2 Hz), 1.55 (1H, ddd, J=4.0, 13.0, 13.0 Hz), 1.62~1.68 (1H, m), 1.74 (1H, ddd, J=3.0, 3.0, 13.0 Hz), 1.93 (1H, br.s, OH), 2.00~2.11 (1H, m), 2.07 (1H, dd, J=5.0, 12.2 Hz), 3.23 (1H, dd, J=11.0, 11.0 Hz), 3.55 (1H, ddd, J=2.0, 4.5, 11.0 Hz), 3.58 (1H, dd, J=11.5, 11.5 Hz), 3.84 (1H, dd, J=5.0, 11.5 Hz), 3.98 (1H, ddd, J=5.0, 11.0, 11.0 Hz), 4.58 (1H, dd, J=7.0, 11.0 Hz), 4.66 (1H, dd, J=3.5, 11.0 Hz), 9.14 (2H, d, J=2.0 Hz), 9.24 (1H, t, J=2.0 Hz). (Found: C, 53.82; H, 5.67; N, 6.63. Calc for C19H2409N2: C, 53.77; H, 5.70; N, 6.60 %).

(3R,4S,6R,9R)-9-Ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro[5,5]undecane [(-)-talaromycin A] 1a and (3S,4S,6R,9R)-9-Ethyl-4-hydroxy-3-hydroxymethyl-1,7-dioxaspiro[5,5]undecane [(-)-talaromycin B] 2a. 1e (100 mg, 0.24 mmol) was treated with K_2OO_3 (36 mg) in MeOH (1 ml)-THF (0.5 ml) at room temp followed by chromatographic purification [SiO₂ (0.8 g), <u>n</u>hexane-EtOAc (4:1~2:1) followed by Et₂0] to give talaromycin A la (57 mg, quantitative). Analytical sample [hygroscopic needles from n-pentane-Et₂O]: m.p. 19-20°; [a] β^4 -146° (c=0.46) [lit.⁸ [a] β^0 -110.2° (c=0.83, CHCl₃), 91-93 & e.e., lit.⁹ [a] β^6 -124.9° (c=1.11, CHCl₃)]; ORD (c=8.9 x 10⁻², MeOH; t=25°) [[a] (λ_1 nm)] -129° (600), -200° (500), -365° (400), -516° (350), -786° (300), -1382° (250); vmax 3420 (s), 2970 (s), 2950 (s), 2920 (s), 2880 (s), 1470 (m), 1450 (m), 1450 (m), 1380 (m), 1340 (w), 1290 (w), 1260 (w), 1250 (w), 1240 (m), 1215 (m), 1185 (s), 1160 (s), 1150 (m), 1125 (w), 1105 (s), 1075 (vs), 1045 (s), 1020 (s), 995 (vs), 970 (m), 940 (w), 925 (w), 890 (w), 860 (s), 805 (m), 770 (m), 760 (m), 695 (w) cm⁻¹. vmax (CHCl₃) 3680 (w), 3620 (s), 3450 (s), 3020 (s), 2980 (s), 2950 (s), 2900 (s), 1465 (s), 1445 (s), 1385 (s), 1330 (m), 1290 (m), 1265 (s), 1180 (s), 1160 (s), 1130 (m), 1100 (vs), 1070 (vs), 1040 (vs), 995 (vs), 955 (m), 945 (m), 930 (w), 910 (w), 885 (w), 865 (s), 830 (w), 670 (s), 585 (m), 570 (m) cm⁻¹. Its IR spectrum as CHCl₃ soln was identical with that provided by Prof. A. B. Smith, III. & (400 MHz) 0.88 (3H, t, J=7.5 Hz), 1.06~1.24 (2H, m), 1.38 (1H, dddd, J-4.0, 13.0, 13.0, 13.0 Hz), 1.36~1.48 (1H, m), 1.51 (1H, ddd, J=4.2, 13.0, 13.0 Hz), 1.59~1.66 (1H, m), 1.67~1.73 (1H, m), 1.72 (1H, dd, J=11.0, 13.0 Hz), 1.89 (1H, ddd, J=1.0, 5.0, 13.0 Hz), 2.11~2.18(1H, m), 2.25 (2H, br.s, OHx2), 3.19 (1H, dd, J=11.0, 11.0 Hz), 3.52 (1H, ddd, J=2.0, 4.5, 11.0 Hz), 3.58 (1H, dd, J=1.5, 12.0 Hz), 3.75 (1H, dd, J=2.8, 12.0 Hz), 3.80 (1H, dd, J=5.0, 11.0 Hz), 4.21 (1H, dd, J=9.0, 11.0 Hz), 4.41 (1H, ddd, J=5.0, 5.5, 11.0 Hz). Its ¹H NMR spectrum was identical with that reported.¹ ¹³C NMR 6 11.1, 24.9, 25.2, 35.3, 37.0, 40.4, 41.3, 60.9, 61.7, 65.4, 67.1, 97.2. The 6 value of the spiro center (97.2) was in good accord with that reported (97.2).¹ $MS^{cf.5b}$: m/z 231 (M⁺+1), 230 (M⁺), 212, 200, 157, 153, 147, 146, 145, 144, 143, 129, 127, 126, 125, HR-MS: m/2 230,1525 (M⁺). Calc for C12H22O4: 230,1518, (Found: C, 61.89; H, 9.65. Calc for $C_{12}H_{22}O_4^{*(H_2O)/9}$: C, 62.06; H, 9.66 %). 1a (54.4 mg) was treated with Amberlyst[®]-15 (10 mg) in MeOH (1 ml) followed by chromatographic purification [SiO₂ (1

g), n-hexane-EtOAc (3:1~2:1) followed by Et₂O] to give 2a (42 mg, 78 % from 1e) as a colorless viscous oil, m_{0}^{55} 1.4828; $[\alpha]_{0}^{24}$ -B9,1° (c=0.48) [1it.⁸ ($\alpha]_{0}^{20}$ -B4,1° (c=0.46, CHCl₃), 91~93 % e.e.]; ORD (c=5.6 x 10⁻², NeOH; t=24°) [[α] (λ , nm)] -357° (600), -393° (500), -473° (400), -508° (350), -625° (300), -848° (250); vmax 3370 (s), 2990 (s), 2950 (s), 2900 (s), 1470 (s), 1450 (s), 1295 (w), 1265 (w), 1245 (w), 1215 (m), 1185 (s), 1160 (s), 1130 (s), 1090 (e), 1070 (s), 1040 (s), 1010 (s), 990 (s), 995 (s), 955 (w), 930 (w), 890 (s), 875 (s), 825 (w), 815 (w), 795 (s), 705 (s), 700 (w), 690 (w) cm⁻¹; vmax (CHCl₃) 3670 (w), 3620 (s), 1240 (s), 1240 (s), 1225 (s), 1180 (s), 1155 (s), 1140 (s), 1125 (s), 1040 (s), 1340 (w), 1330 (w), 1295 (m), 1260 (s), 1240 (s), 1225 (s), 1180 (s), 1155 (s), 1140 (s), 1125 (s), 1085 (s), 1070 (s), 1040 (s), 1010 (vs), 990 (s), 970 (m), 955 (m), 930 (m), 890 (s), 870 (s), 665 (w), 615 (m), 575 (m), 565 (m). Its IR spectrum as CHCl₃ soln was virtually identical with that provided by Prof. A. B. Smith, III. & (400 MHz) 0.488 (3H, t, J=7.5 Hz), 1.06-1.24 (2H, m), 1.40 (1H, dddd, J=3.6, 13.0, 13.0, 13.0, 12.0, 14.5 (1H, dd., J=11.0, 11.0, 12.5 Hz), 1.39~1.50 (1H, m), 1.79 (1H, ddd, J=2.5, 3.6, 13.0 Hz), 1.78~1.91 (1H, m), 1.99 (1H, dd, J=2.0, 4.5, 11.0, Hz), 3.59 (1H, dd, J=5.0, 11.0 Hz), 3.70 (2H, d, J=6.0 Hz), 4.05 (1H, ddd, J=5.0, 10.5, 11.0 Hz), 3.51 (1H, ddd, J=2.0, 4.5, 11.0 Hz), 3.59 (1H, dd, J=5.0, 11.0 Hz), 3.70 (2H, d, J=6.0 Hz), 4.05 (1H, ddd, J=5.0, 10.5, 11.0 Hz), 3.51 (1H, ddd, J=2.5, 65.3, 67.2, 97.2. Its ¹³C NMR spectrum was identical with that of (t)-2a reported previously.⁵⁵ MS⁵E: m/z 231 (M⁺1), 230 (M⁺), 212, 200, 157, 153, 147, 146, 145, 144, 143, 129, 127, 126, 125. HR-MS: m/z 230.1525 (M⁺). Calc for C₁₂H₂Z₄: 230.1518.

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